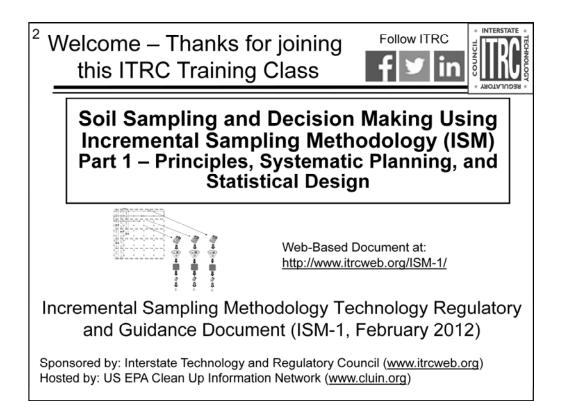


No associated notes.



When sampling soil at potentially contaminated sites, the goal is collecting representative samples which will lead to quality decisions. Unfortunately traditional soil sampling methods don't always provide the accurate, reproducible, and defensible data needed. Incremental Sampling Methodology (ISM) can help with this soil sampling challenge. ISM is a structured composite sampling and processing protocol that reduces data variability and provides a reasonable estimate of a chemical's mean concentration for the volume of soil being sampled. The three key components of ISM are systematic planning, field sample collection, and laboratory processing and analysis. The adequacy of ISM sample support (sample mass) reduces sampling and laboratory errors, and the ISM strategy improves the reliability and defensibility of sampling data by reducing data variability.

ISM provides representative samples of specific soil volumes defined as Decision Units. An ISM replicate sample is established by collecting numerous increments of soil (typically 30 to 100 increments) that are combined, processed, and subsampled according to specific protocols. ISM is increasingly being used for sampling soils at hazardous waste sites and on suspected contaminated lands. Proponents have found that the coverage afforded by collecting many increments, together with disciplined processing and subsampling of the combined increments, yields consistent and reproducible results that in most instances have been preferable to the results obtained by more traditional (e.g. discrete) sampling approaches.

This 2-part training course along with ITRC's web-based Incremental Sampling Methodology Technical and Regulatory Guidance Document (ISM-1, 2012) is intended to assist regulators and practitioners with the understanding the fundamental concepts of soil/contaminant heterogeneity, representative sampling, sampling/laboratory error and how ISM addresses these concepts. Through this training course you should learn:

- basic principles to improve soil sampling results

- systematic planning steps important to ISM
- how to determine ISM Decision Units (DU)
- the answers to common questions about ISM sampling design and data analysis
- methods to collect and analyze ISM soil samples
- the impact of laboratory processing on soil samples
- how to evaluate ISM data and make decisions

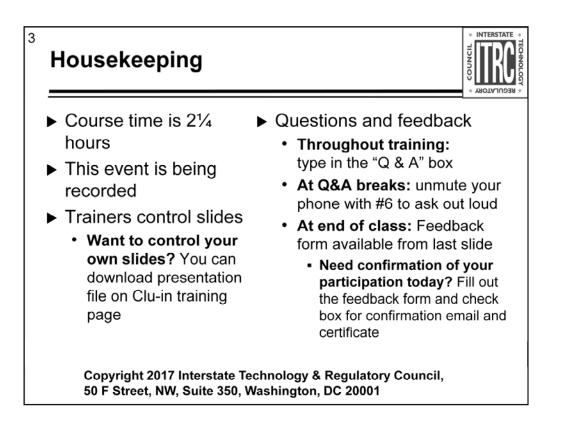
In addition this ISM training and guidance provides insight on when and how to apply ISM at a contaminated site, and will aid in developing or reviewing project documents incorporating ISM (e.g., work plans, sampling plans, reports). You will also be provided with links to additional resources related to ISM.

The intended users of this guidance and training course are state and federal regulators, project managers, and consultant personnel responsible for and/or directly involved in developing, identifying or applying soil and sediment sampling approaches and establishing sampling objectives and methods. In addition, data end users and decision makers will gain insight to the use and impacts of ISM for soil sampling for potentially contaminated sites.

Recommended Reading: We encourage participants to review the ITRC ISM document (<u>http://www.itrcweb.org/ISM-1/</u>) prior to participating in the training classes. If your time is limited in reviewing the document in advance, we suggest you prioritize your time by reading the Executive Summary, Chapter 4 "Statistical Sampling Designs for ISM," and Chapter 7 "Making Decisions Using ISM Data" to maximize your learning experience during the upcoming training classes.

ITRC (Interstate Technology and Regulatory Council) www.itrcweb.org

Training Co-Sponsored by: US EPA Technology Innovation and Field Services Division (TIFSD) (<u>www.clu-in.org</u>) ITRC Training Program: training@itrcweb.org; Phone: 402-201-2419

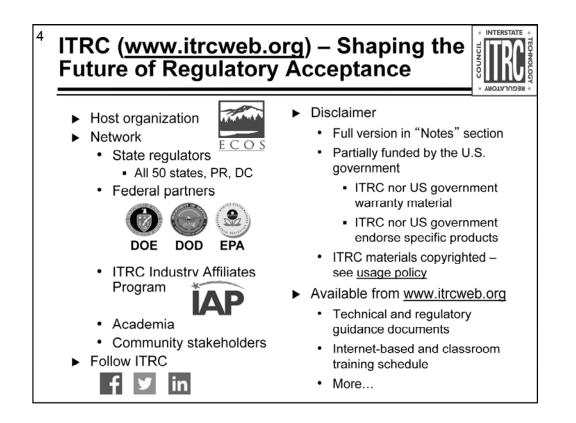


Although I'm sure that some of you are familiar with these rules from previous CLU-IN events, let's run through them quickly for our new participants.

We have started the seminar with all phone lines muted to prevent background noise. Please keep your phone lines muted during the seminar to minimize disruption and background noise. During the question and answer break, press #6 to unmute your lines to ask a question (note: *6 to mute again). Also, please do NOT put this call on hold as this may bring unwanted background music over the lines and interrupt the seminar.

Use the "Q&A" box to ask questions, make comments, or report technical problems any time. For questions and comments provided out loud, please hold until the designated Q&A breaks.

Everyone – please complete the feedback form before you leave the training website. Link to feedback form is available on last slide.



The Interstate Technology and Regulatory Council (ITRC) is a state-led coalition of regulators, industry experts, citizen stakeholders, academia and federal partners that work to achieve regulatory acceptance of environmental technologies and innovative approaches. ITRC consists of all 50 states (and Puerto Rico and the District of Columbia) that work to break down barriers and reduce compliance costs, making it easier to use new technologies and helping states maximize resources. ITRC brings together a diverse mix of environmental experts and stakeholders from both the public and private sectors to broaden and deepen technical knowledge and advance the regulatory acceptance of environmental technologies. Together, we're building the environmental community's ability to expedite quality decision making while protecting human health and the environment. With our network of organizations and individuals throughout the environmental community, ITRC is a unique catalyst for dialogue between regulators and the regulated community.

For a state to be a member of ITRC their environmental agency must designate a State Point of Contact. To find out who your State POC is check out the "contacts" section at www.itrcweb.org. Also, click on "membership" to learn how you can become a member of an ITRC Technical Team.

Disclaimer: This material was prepared as an account of work sponsored by an agency of the United States Government. Neither the United States Government nor any agency thereof, nor any of their employees, makes any warranty, express or implied, or assumes any legal liability or responsibility for the accuracy, completeness, or usefulness of any information, apparatus, product, or process disclosed, or represents that its use would not infringe privately owned rights. Reference herein to any specific commercial product, process, or service by trade name, trademark, manufacturer, or otherwise does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States Government or any agency thereof. The views and opinions of authors expressed herein do not necessarily state or reflect those of the United States Government or any agency thereof and no official endorsement should be inferred.

The information provided in documents, training curricula, and other print or electronic materials created by the Interstate Technology and Regulatory Council ("ITRC" and such materials are referred to as "ITRC Materials") is intended as a general reference to help regulators and others develop a consistent approach to their evaluation, regulatory approval, and deployment of environmental technologies. The information in ITRC Materials was formulated to be reliable and accurate. However, the information is provided "as is" and use of this information is at the users' own risk.

ITRC Materials do not necessarily address all applicable health and safety risks and precautions with respect to particular materials, conditions, or procedures in specific applications of any technology. Consequently, ITRC recommends consulting applicable standards, laws, regulations, suppliers of materials, and material safety data sheets for information concerning safety and health risks and precautions and compliance with then-applicable laws and regulations. ITRC, ERIS and ECOS shall not be liable in the event of any conflict between information in ITRC Materials and such laws, regulations, and/or other ordinances. The content in ITRC Materials may be revised or withdrawn at any time without prior notice.

ITRC, ERIS, and ECOS make no representations or warranties, express or implied, with respect to information in ITRC Materials and specifically disclaim all warranties to the fullest extent permitted by law (including, but not limited to, merchantability or fitness for a particular purpose). ITRC, ERIS, and ECOS will not accept liability for damages of any kind that result from acting upon or using this information.

ITRC, ERIS, and ECOS do not endorse or recommend the use of specific technology or technology provider through ITRC Materials. Reference to technologies, products, or services offered by other parties does not constitute a guarantee by ITRC, ERIS, and ECOS of the quality or value of those technologies, products, or services. Information in ITRC Materials is for general reference only; it should not be construed as definitive guidance for any specific site and is not a substitute for consultation with qualified professional advisors.

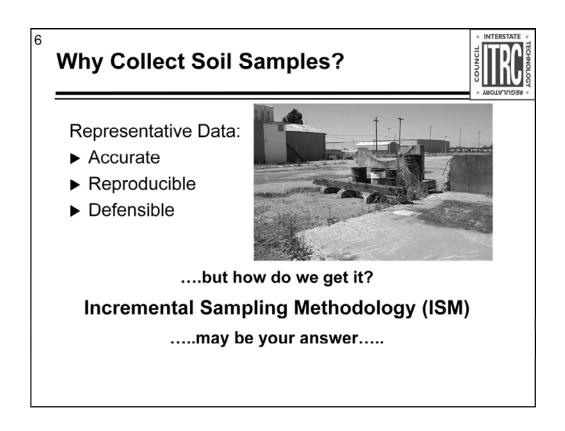


Annette C. Dietz, Ph.D., has been an instructor in the Department of Civil and Environmental Engineering at Portland State University in Portland, Oregon since 2015. From 2010 to 2015, she was the Cleanup Program Coordinator for the Oregon Department of Environmental Quality (OR DEQ) located in Portland, Oregon. She was a technical and policy expert for cleanup program activities, oversaw the maintenance and development of policy and guidance for cleanup project work, coordinated program functions performed by technical staff in regional offices, and provided outreach and training support for external stakeholders. Before joining OR DEQ in 2010, Annette worked as an environmental consultant managing and performing site investigation and remediation projects. While at OR DEQ, she was Oregon's Point of Contact (POC) to ITRC and a member of the ITRC Incremental Sampling Methodology team. Annette earned a bachelor's degree in Spanish and Global Studies in 1993, a master's in environmental engineering in 1996 and a doctoral degree in environmental engineering in 2000, all from the University of Iowa in Iowa City, Iowa.

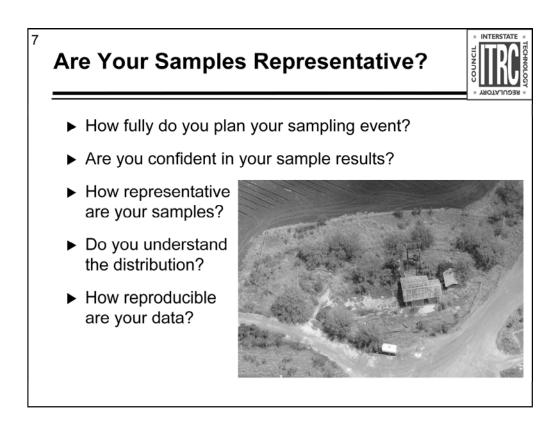
Deana Crumbling is an Environmental Scientist in the Technology Innovation section of EPA's Office of Solid Waste and Emergency Response in Washington, DC. Since 1997 she has focused on the topics of dynamic work plans, field analysis and other analytical chemistry technologies, sampling designs and data uncertainty management. She has taught many classroom and webinar courses in those topics. Before coming to EPA, she worked for 2 years as a risk assessor for an environmental consulting firm, for 2 years as a lab and safety manager and adjunct faculty for a college, for 1 year as a science consultant for an environmental attorney, and 1 year in the Pennsylvania State Hazardous Site Cleanup Program. She worked for 20 years as a hospital-based clinical laboratory analyst and trainer. Deana was a team member in the ITRC Site Characterization and Monitoring Team from 2002 to 2004, and now a member of the Incremental Sampling Methodology Team since 2009. She earned a bachelor's in biochemistry and a bachelor's in psychology from Lebanon Valley College in Lebanon, Pennsylvania, in 1989, and a master's in environmental science from Drexel University in Philadelphia, Pennsylvania, in 1997.

Robin Boyd is a Senior Project Manager with AECOM in Virginia. Robin is a specialist in the use of incremental sampling for both surface and subsurface remedial investigations and remediation. Since 1989 Robin has gained experience with investigating hazardous waste sites including the development and implementation of innovative technologies for the remediation of soil and ground water. He has designed and implemented over 35 incremental sampling programs of various types and conducted numerous training classes on the use of incremental sampling. He has attended both Francis Pitard's and Chuck Ramsey's classes on incremental sampling and Gy's Sampling Theory (which is the cornerstone of ISM). He is an active member of the ITRC Incremental Sampling Methodology team. Robin earned both a bachelor's degree in 1979 and master's degree in 1981 in geology and geophysics from the University of Wisconsin-Madison. Robin is also a registered professional geologist.

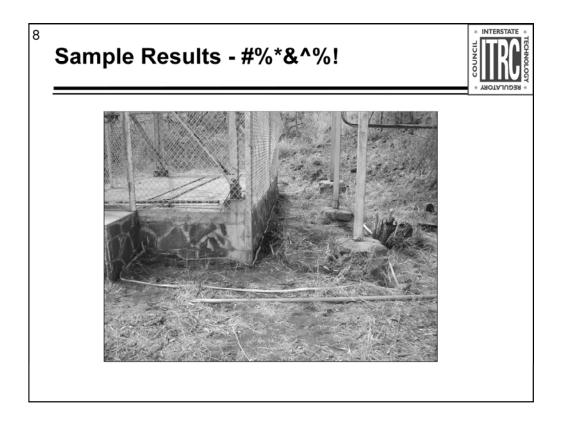
Philip Goodrum is a Senior Managing Scientist with Integral Consulting in Syracuse, New York. Since 1989 Phil has gained experience in quantitative risk assessment and environmental modeling, specializing in applications to human health and ecological risk assessment, sediment remediation, groundwater compliance monitoring, and natural resource damage assessment. He brings a broad understanding of the use and effective communication of data evaluation, visualization, and statistical analysis techniques to support ecological risk assessment and injury assessment. He is responsible for developing sampling designs and conducting data interpretation, statistical analysis, modeling, and risk characterization at sites around the country. He is a recognized national expert in probabilistic modeling, lead (Pb) risk assessment, and environmental sampling, having been invited to teach numerous professional short courses on these topics by regulators and industry. He has co-authored USEPA guidance on probabilistic risk analysis, served as an independent peer reviewer for state and federal agencies on survey design, and continues to serve on USEPA's Science Advisory Board for lead (Pb). Phil has contributed to ITRC Incremental Sampling Methodology team since 2009. Phil earned his bachelor's degree in environmental technology from Cornell University in Ithaca, New York in 1989, a master's in water resources from SUNY Environmental Science and Forestry (ESF) in Syracuse, New York in 1995, and Ph.D. in environmental engineering from ESF in 1999. He is on the adjunct faculty at Syracuse University where he teaches a course in environmental toxicology.



Picture Reference: http://www.swrcb.ca.gov/rwqcb2/brownfields.shtml



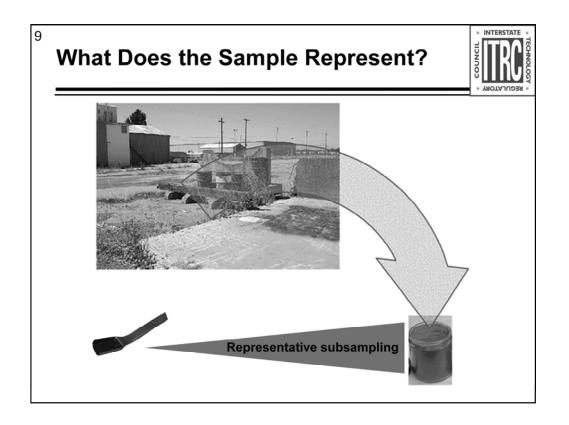
How are your decisions made when you review soil sampling data?



Discrete samples – if you find a hotspot you step out... but if you get a ND – are you done?

With a small number of discrete samples how well did you define the extent of contamination to begin with?

What does each discrete sample represent?



What does a single sample represent and how can/should you assess the spatial variation of samples.

A small # of discrete samples encourages two key errors:

- 1) Underestimate the representative concentration in the an area, and
- 2) Underestimate the vertical and later extent of contamination.

Does 1g of soil represent your site?

That small volume of samples that is analyzed provides a results that represents the area we are assessing for risk type decisions.

Picture Reference: http://www.swrcb.ca.gov/rwqcb2/brownfields.shtml

¹⁰ What Do These Environmental Criteria Have In Common?



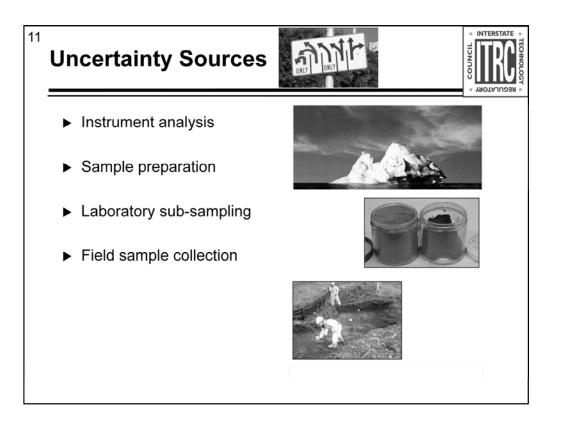
- Most risk-based environmental criteria based on estimate of mean
 - · Soil screening levels
 - Regional screening levels
 - Site-specific cleanup levels
 - · Exposure point concentrations

Most Risk-based environmental criteria are based on an estimate of the mean (.e.g., 95% UCL).

If you have a few discrete samples how do you estimate the mean?

Discrete samples give some sense of spatial variability.

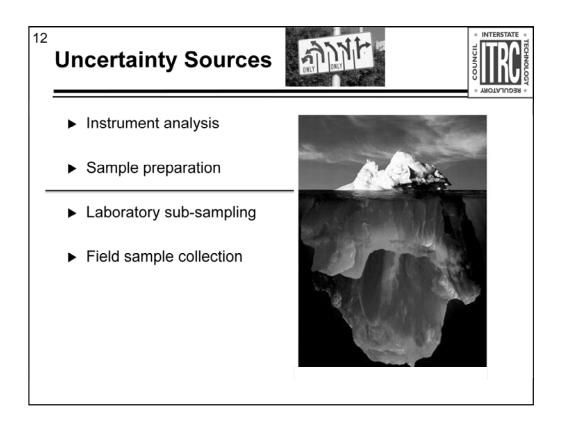
The more discrete samples you have, the more you know about the population variability, and build certainty.



What are Sources of Uncertainty and where do we find them?

With proper maintenance and standard operating procedures instrument analysis is actually very consistent.

Uncertainty increases as you move down the list.



The largest amount of uncertainty lies in laboratory sub sampling and field sampling collection, including sample heterogeneity.

Is there a better way to control uncertainty and errors and get a better representative sample?

³ What is Incremental Sampling Methodology (ISM)?



ISM Objective: To obtain a single sample for analysis that has the mean analyte concentration representative of the decision unit

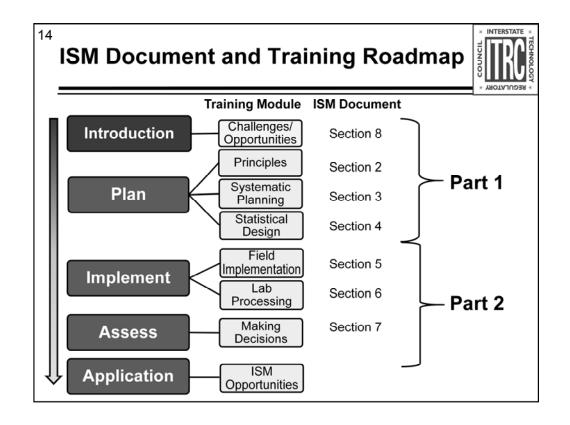
- Structured composite sampling and processing protocol
- Reduces data variability
- Provides a reasonably unbiased estimate of mean contaminant concentrations in a volume of soil targeted for sampling

Decision Unit (DU): the smallest volume of soil (or other media) for which a decision will be made based upon ISM sampling

Incremental Sampling is a structured sampling and processing protocol that reduces data variability and increases sample representativeness.

ISM is an improved form of composite sampling, because the process involved in collection and analysis of an ISM sample greatly improves sample representativeness. The ISM sample goal is to have all the same constituents in the same proportion as the volume sampled.

ISM samples provide a result that better estimates the mean concentration of the sampled volume, than sparse discrete sampling.



The Internet-based training follows the ISM document section by section

Two-part ITRC Internet-based training:

Part 1 - Principles, Systematic Planning, and Statistical Design

Part 2 - Field Implementation, Lab processing, and Data Assessment

The planning, field implementation, and laboratory processing are critical to collecting a sample that yields Highly Reproducible Mean Concentrations

¹⁵ 2009 ISM Survey: Areas of Question/Concern



263 responses (75% respondents state regulators and consultants)

- ► Can ISM find "hot spots"?
- ► Do regulators allow or accept ISM?
- Can you collect volatile organic compound (VOC) samples?
- Does ISM delineate the extent of contamination?
- ▶ What's the right size for a Decision Units (DUs)?
- ► Can you obtain Upper Confidence Limits (UCLs)?
- Can ISM and discrete results be compared?
- ► Are there approved laboratory processes and certification?
- ► How much does ISM cost?

Reluctance to use ISM stems from a lack of experience

ITRC, ISM-1, Appendix B, August 2009 ISM Survey Results

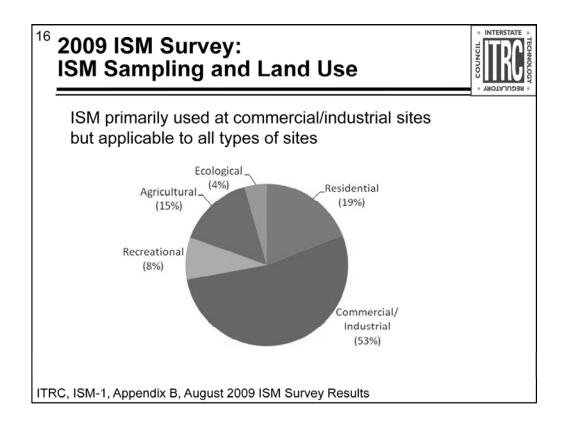
Survey identified outstanding issues and used to aid development of the ISM technical regulatory document.

263 responses to our survey
½ of the responses from consultants
¼ from State regulators
¼ from federal agencies, regulators, laboratories, stakeholder groups.

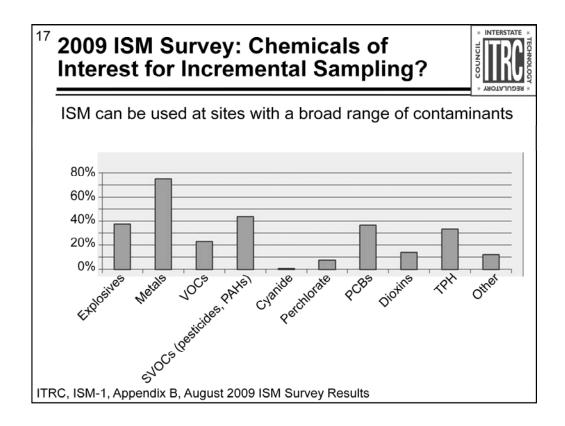
These summarize the key implementation issues identified by the survey.

Section 8 addressed these issues in the ISM document.

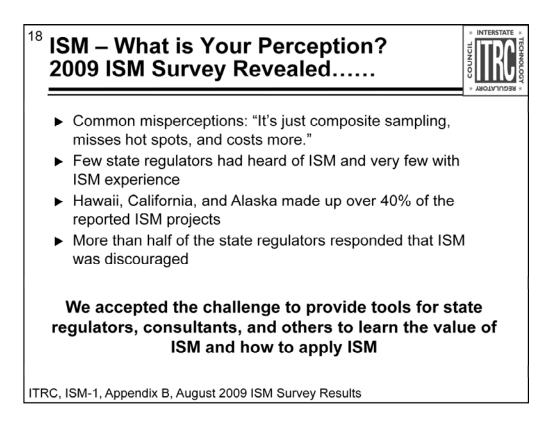
ISM is still being developed and some of these issues (e.g. cost) are still being worked out with more wide spread use.



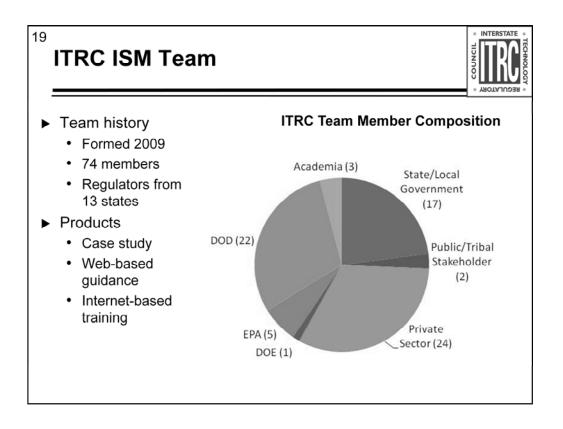
The survey found that ISM has been applied on a variety of sites.



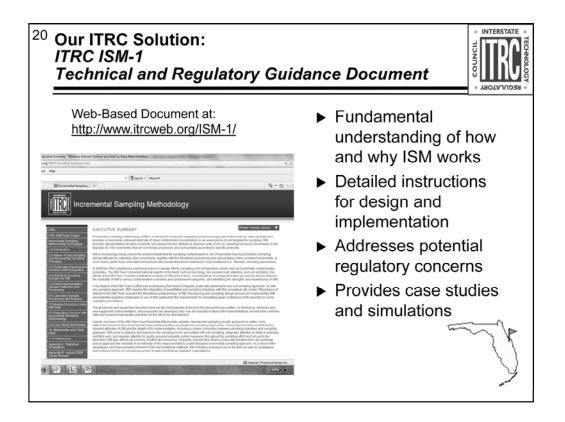
ISM sampling can be used for a broad range of contaminants - data from our 2009 survey.



No associated notes



Multi-disciplinary team includes, scientists, geologist, toxicologists, engineers, chemists, statisticians, and community and tribal representatives.



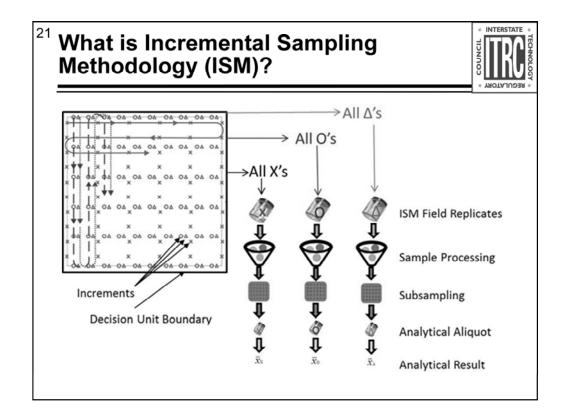
Training classes on Multi-Increment sampling and guidance available from Hawai'i, Alaska, and the USACOE

ISM Tech-Reg document deals with things in more detail and include case study and simulation info to support application of ISM.

In 2010, the ISM Team developed and implement an IS plan for a case study site in Florida. It is presented in our Tech-Reg document and will be presented during different Internetbased training modules.

As of March 2012, the Florida Dept. of Environmental Protection is in the process of developing ISM guidance by incorporating the ITRC ISM guidance document.

Web-based document has live links and you can print the entire document, a Section, or just a page.



ISM involves planning, field implementation, and laboratory processing and subsampling to provide a representative analytical aliquot. The goal of ISM is to analyze an aliquot that represents the same proportion of constituents as the sampling area.

The box represents a Decision Unit or DU - the volume being sampled and the volume you want to make a decision on.

Within the DU, there are sample locations or "Increments".

The sampling grid and increment locations are established during the systematic planning as are the number of increments represented by the X's, O's and triangles.

The ISM field replicate for X is a composite of all 60 X increments, likewise for the O's and the triangles.

After collection in the field the samples are sent to a lab to be processed, and subsampled.

The lab subsampling approach is similar to that applied to collection of a single field increments. The goal of ISM is to have all the same constituents in the same proportion as the area sampled.

The ISM document recommends at least three replicate results (X, O, and triangle) for each DU.

Advantages and Limitations of ISM

22



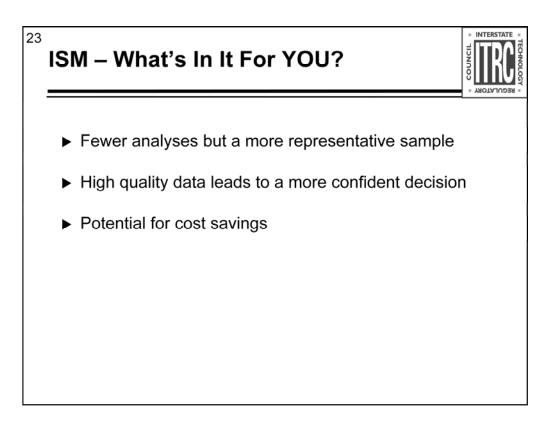
Advantages of ISM	Effect
Improved spatial coverage (increments x replicates)	 Sample includes high and low concentrations in proper proportions
Higher Sample Mass	 Reduces errors associated with sample processing and analysis
Optimized processing	Representative subsamples for analysis
Fewer non-detects	Simplifies statistical analysis
More consistent data	More confident decision
Limitations of ISM	Effect
Small number of replicates	 Limits Upper Confidence Limit calculation methods
No spatial resolution within Decision Unit	Limits remediation options within Decision UnitLimits multivariate comparisons
Assessing Acute Toxicity	Decision Unit has to be very small

ISM has both advantages and disadvantages from a sampling design perspective.

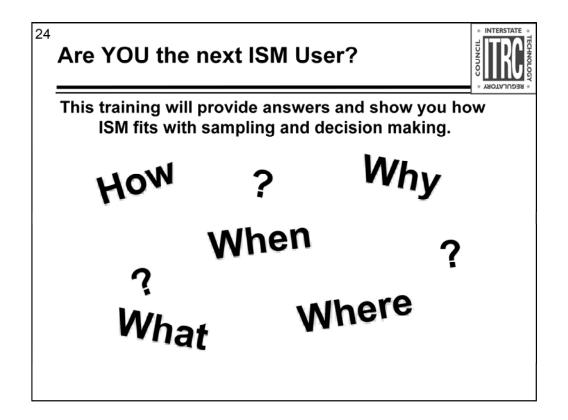
Can't directly compare discrete and ISM samples because each measure different properties of the population.

Under disadvantages, discrete sampling allows for calculations of ratios of two variables – allows for correlations among constituents, or estimates of bioaccumulation factors (update from abiotic media to organisms) that you cannot get from ISM.

When assessing acute toxicity issues, the decision unit would have to be very small for incremental sampling. ISM may not be practical.

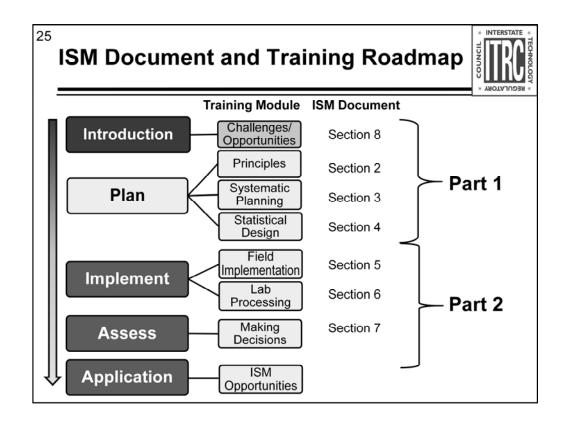


No associated notes



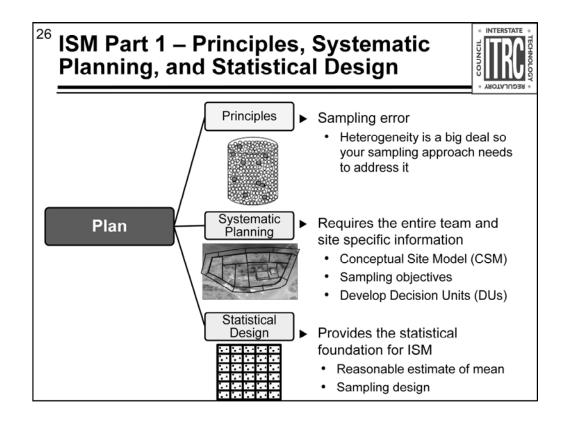
Training provides answers to make informed ISM decisions by answering:

Where can ISM be used? When should ISM <u>not</u> be used? What contaminants are most suitable for ISM? What effect does sample processing have on contaminant concentration? Does ISM mask area of high concentrations due to compositing and homogenization? How does ISM differ from discrete sampling? How many replicates should be collected? How are data quality objectives (DQOs) addressed? How do ISM results relate to action levels?



Part 1 includes Modules on:

Soil Principles, Systematic Planning, and Statistical Design



Soil Principles - Be aware of issues related to heterogeneity and sampling errors

Systematic Planning - Involve the entire team, regulators, consultants, responsible parties in critical elements (e.g. conceptual site model, establish sampling objectives and decision units.) Sampling objectives should drive your sampling design, and the scale of decision making should align with sampling objectives.

Statistical Design - Provides the statistical foundation and describes why ISM provides a reasonable mean, describes a good ISM sampling design, and informs you how ISM provides 95% UCL.





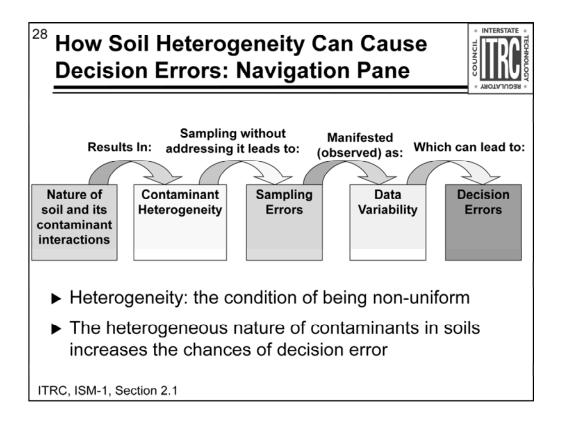
Learn how to use basic principles to improve planning, implementation and decision-making:

- Soil heterogeneity at 2 spatial scales makes it difficult to correctly interpret data results
 - Those spatial scales are micro-scale and short-scale
 - Heterogeneity at these scales can cause data variability → costly decision errors
- Micro-scale heterogeneity is managed by increasing sample mass and improving lab sample processing (required by ISM)
- Short-scale spatial heterogeneity is managed by the field incremental sampling of ISM

ITRC, ISM-1, Sections 2 and 5.3.1

Speaker Notes

• Micro-scale heterogeneity is also called compositional heterogeneity.



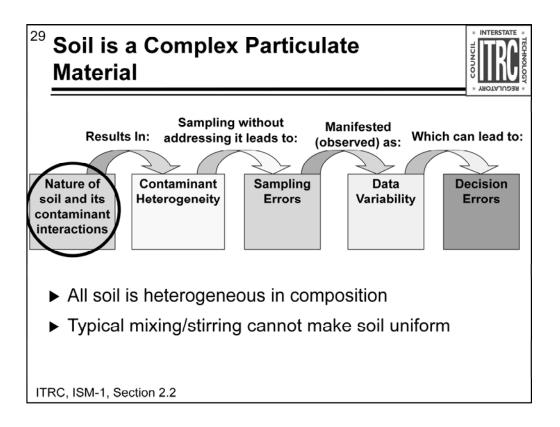
Previous section emphasized 1) importance of <u>up-front planning BEFORE</u> going to the field, and 2) sample processing is critical part of incremental sampling methodology. Planning is necessary to minimize errors throughout data generation. In order to plan effectively, need to understand the causes and effects of sampling error, and be able to detect when sampling error may have degraded data quality.

A key goal of ISM is to reduce decision errors when dealing with soils. Decision error refers to decisions that would be made differently IF the true nature of the contamination were known. An example of a decision error is deciding that contamination is not present above a certain level, when it actually is.

To reduce decision errors, need to examine their root causes. This slide shows how the heterogeneous nature of soil ends up causing decision errors. Starting with the first box on the left (the blue box), we'll look at how contaminants interact with soil. That interaction leads to contaminant heterogeneity, which is a primary cause of sampling errors. Sampling errors, in turn, lead to data variability, and data variability can mislead decision makers. In this presentation I'm going to explain how this cascade occurs. We'll use this figure to serve as a navigation aid.

Supplemental Information

See ISM-1 Section 2.1

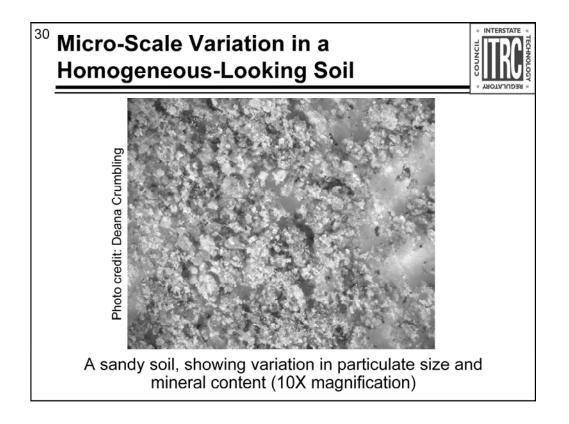


• Heterogeneity in composition is referred to as "compositional" or "constitutional" heterogeneity

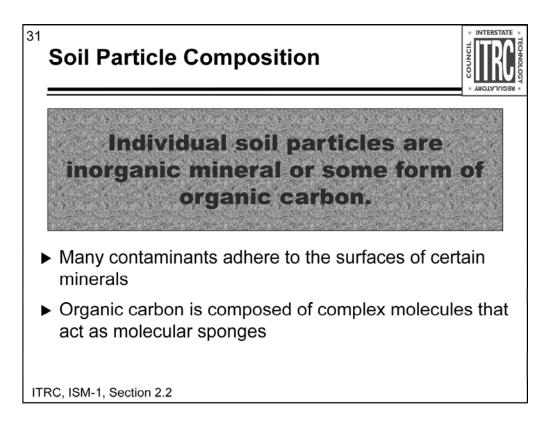
- Soil samples show compositional heterogeneity in 3 primary ways:
 - made up of particle <u>sizes</u> that vary over several orders of magnitude and so differ in surface area, and
 - composed of different <u>mineral</u> grains which vary in their "stickiness" for contaminant molecules,
 - contain various amounts and types of organic matter.

Supplemental Information

See ISM-1 Section 2.2



- This figure is a photo of magnified sandy soil containing very little organic matter.
- At the macro level (i.e., viewed without magnification) this soil looks homogeneous.
- But under magnification, can see it is composed of mineral grains that vary from relatively large to barely visible at this magnification.
- Colors of individual grains vary from white to pink to greenish, reflecting the different minerals present in the grains.
- This is an example of soil heterogeneity at the micro-scale.

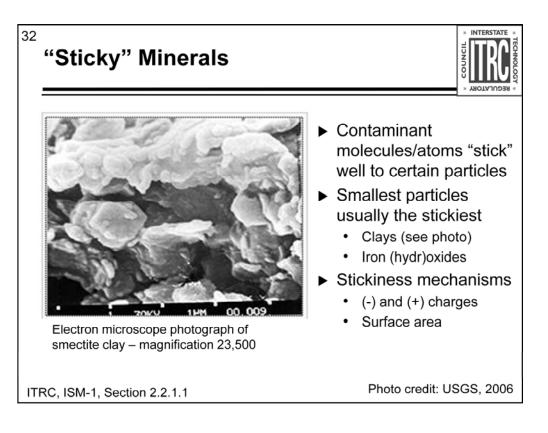


• Soil is composed of an infinite number of particles, some made of inorganic minerals and some of organic matter.

- Organic matter comes from living organisms. It could be grass, leaves, sticks, insects, microbes, etc.
- Organic matter is of particular importance to contaminants because it decays into complex molecules that act as molecular sponges.
- Many contaminants, although not all, adhere well to certain soil minerals.
- Organic and inorganic contaminants can be absorbed into organic carbon complexes.

Supplemental Information

See ISM-1 Section 2.2



• Different particles bind contaminants to varying degrees.

• Clay minerals strongly bind many contaminants.

• This is a photograph of clay particles under high magnification. If you look closely, you'll see that clay particles take the form of stacked plates.

• The plates have molecule-sized spaces between them. The very small size of clay particle gives them a large surface area on the outside, then the plate structure provides even more. More surface area provides more sites where contaminants can bind.

• Some of the "stickiness" of clays is due having many negative charges lining the inside of the plates. Clay particles have some positive charges along the edges of the plates.

• Negative charges attract metal contaminants that are positively charged. An example is when Pb in bullets corrodes into Pb minerals that then dissolve and release positively charged Pb ions into the soil. Lead ions are attracted to the clay's negative charges and can become trapped between the clay plates.

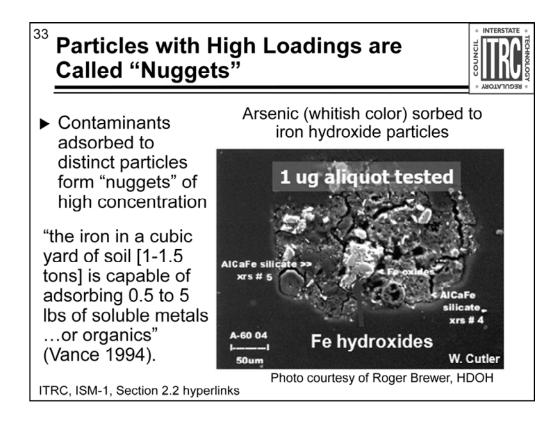
• Another type of soil particle that is sticky from a contaminant's point of view are oxide minerals, such as iron oxides and aluminum oxides. Geochemical oxides are small particles with a large surface area. They can carry either a positive or negative charge depending on the pH.

• Iron oxide is interchangeable with iron hydroxide, depending on the pH.

Supplemental Information

See ISM-1 Section 2.2.1.1

Photo credit: USGS Photo Library, 2006, USGS, URL = http://libraryphoto.cr.usgs.gov/cgibin/show_picture.cgi?ID=ID.%20McKee,%20E.D.%20%20316

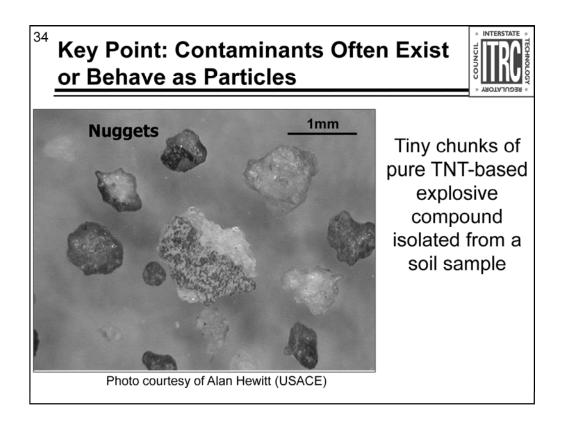


- Particulate iron minerals, such as oxides, are very good at binding contaminants.
- One researcher stated that the Fe in a cubic yard of soil can adsorb ½ to 5 lbs of soluble metals or organics.
- The photomicrograph shows microscopic iron hydroxide grains coated with arsenic. The arsenic appears as a light-colored deposit covering Fe-OH grains (see red arrow).
- Silicate minerals make up most of the soil mass in the photo. Arsenic does not adsorb to those minerals, so they are dark gray.
- Photo provided by Roger Brewer with the Hawaii Dept. of Health

Supplemental Information

See ISM-1 Section 2.2 hyperlinks

Quote from the journal article: "Given the average concentration in soil, the iron in a cubic yard of soil is capable of adsorbing from 0.5 to 5 pounds of soluble metals as cations, anionic complexes, or a similar amount of organic[s]." (Vance, 1994). [Reference = David B. Vance. "Iron – The Environmental Impact of a Universal Element," *National Environmental Journal*, May/June. 1994 Vol.4 No. 3 page 24-25. see also URL = http://2the4.net/iron.htm]



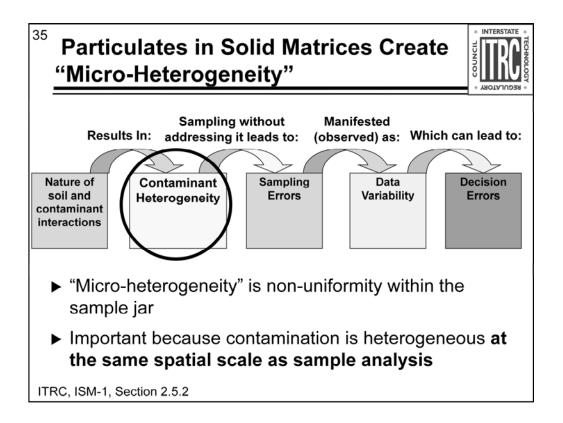
• The photo is of Composition B particles from low-order 81-mm mortar. 60% Military grade RDX (Contains about 10% HMX) 39% Military grade TNT.

• Photo provided by Alan Hewitt (US Army Corp of Engineers Cold Regions Research and Engineering Laboratory).

• Sometimes contaminants are released directly in particulate form. Examples are explosives residues, organic or metal-based pesticides applied as a dust; airborne smelter residues depositing as dust; and lead and other metals dust and fragments created by firing guns at firing ranges.

• But even if they were not originally released in particulate form, contaminants behave as if they were particles when they bind to soil particles by the mechanisms just described.

• As a consequence <u>contaminants are heterogeneous in their spatial distribution</u> throughout even small soil samples.



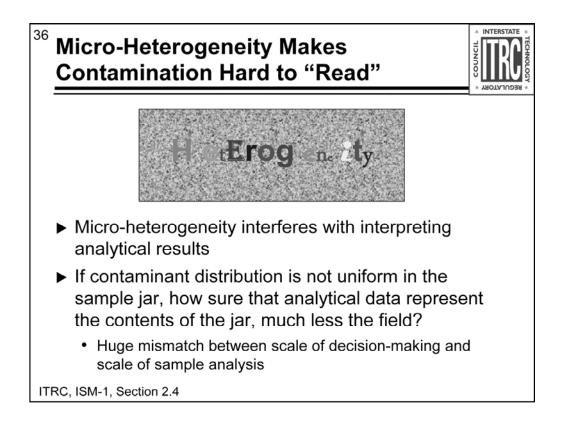
• This is important because particulate contaminants integrate with soil particles in a nonuniform manner, which creates contaminant heterogeneity at a micro-scale. In other words, contaminants are not uniformly spread out evenly throughout the soil in a jar.

• This is called "distributional heterogeneity" at a micro-scale.

• This matters because the mechanics of sample analysis take place at this micro spatial scale. When the lab scoops out subsamples from a jar for analysis, different scoops of soil may have different numbers of contaminant particles.

Supplemental Information

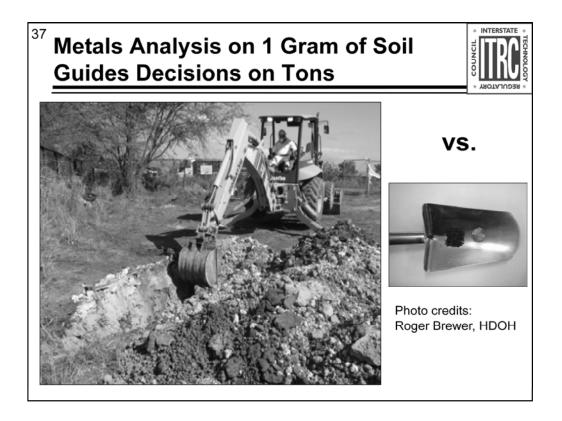
See ISM-1 Section 2.5.2



- Like in this graphic, heterogeneity makes soil contamination hard to read.
- Soil may <u>appear</u> to be homogeneous when viewed from the spatial scale of decisionmaking, but it is NOT homogeneous at the scale at which chemical data are generated.
- Yet we expect that analyzing tiny samples will tell us the true concentration of tons of soil.

Supplemental Information

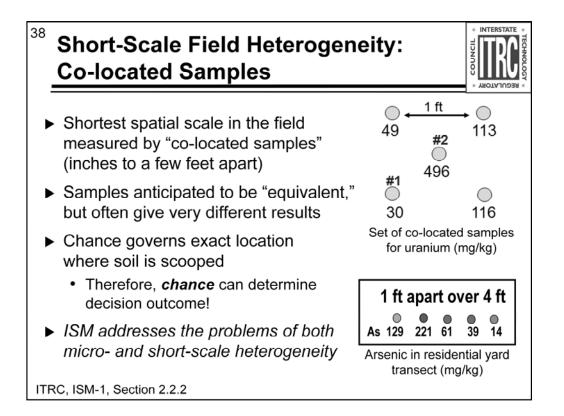
See ISM-1 Section 2.4



• Based on the results of analyses performed on a few grams of soil, decisions are made about whether contamination is present (and at what level) in tens to hundreds to thousands of tons of soil.

• Although a jar of soil containing 100 or more grams of soil is submitted to the lab, routine metals analysis actually analyzes only 0.5, 1 or sometimes 2 gram of soil (depending on the lab) from that jar.

• Organics analysis typically will analyze from 5 to 30 grams (depending on the lab and the analyte).



Previous discussion focused on heterogeneity WITHIN a sample (i.e., within a single jar).

Now will focus on heterogeneity BETWEEN samples (i.e., from one sample to another in the field).

"Co-located samples" are a QC check designed to assess short-scale heterogeneity.

"Co-located samples" are expected to be equivalent in that they are expected to have pretty much the same concentration.

But often co-located samples have very different results even though they may be only inches apart.

Within the confines of a small area, chance governs which spoonful of core of soil is picked to be put in the jar.

Since the concentration of each spoonful might be very different, chance can determine which decision gets made on that area.

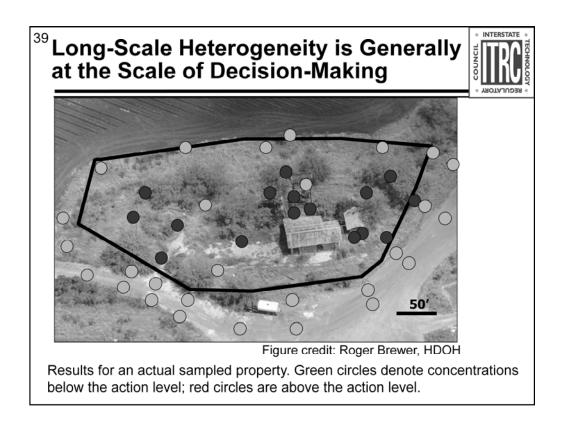
This is one of the dangers of making decisions based on single grab sample results.

Uranium data set: although only about 8 inches apart, sample #1 has a concentration of 30, while sample #2 is nearly 500. If discrete samples are used to make decisions, the decisions in this area would be determined by chance, because the result depends on where the sampler happens to kneel down and dig.

Co-located sample results are affected by both short-scale heterogeneity AND within-sample heterogeneity. So unless you have controlled for within-sample heterogeneity, you won't be able to measure between-sample heterogeneity.

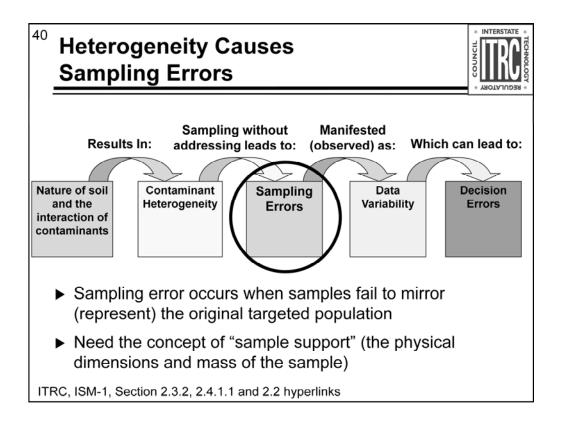
Uranium data source: Robert Johnson (US Dept of Energy) Arsenic data source: Deana Crumbling (USEPA). Data from center of a residential yard.

Supplemental Information See ISM-1 Section 2.2.2



• Long-scale heterogeneity is the spatial scale at which differences in concentration are expected.

• Sampling programs are generally designed to search for variation in concentrations at this scale.



• The next concept is that insufficient control over heterogeneity's effects can lead to sampling errors. A sampling error is said to occur when the sampling process produces a sample that does not represent the intended population.

• To discuss sampling errors, need the term, "sample support."

• <u>Sample Support</u>: "the size, shape, and orientation of sampling volume (i.e., "support") for heterogeneous media have a significant effect on reported measurement values." EPA Soil Screening Guidance: Technical Background Document, EPA/540/R95/128, May 1996

• Sample support refers to the physical dimensions of a sample as it is collected from the parent material.

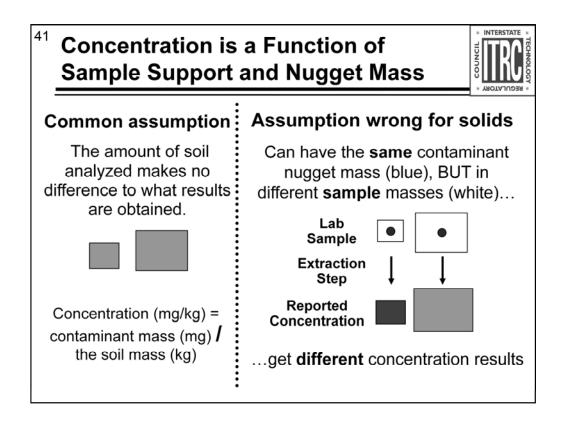
• For example, a 6-inch deep core is a different sample support than scraping up surface soil to a 2-inch depth. A core with a 2-inch diameter is a different sample support from a core with a 5-inch diameter. A 100-gram sample is a different sample support from a 300-gram sample.

Sample support is a critical factor governing the results of soil measurements.

• The following slides explain how this works.

Supplemental Information

See ISM-1 Sections 2.3.2, 2.4.1.1 and 2.2 hyperlinks



• The issue of "sample support" for heterogeneous environmental and waste matrices invalidates the common assumption that the reported concentration of an environmental sample should be the same no matter what mass/volume of sample is collected and analyzed.

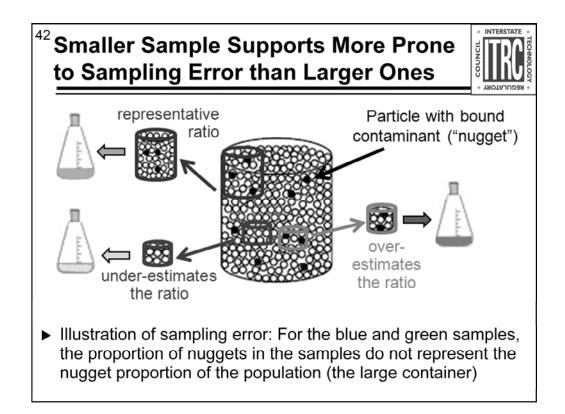
• The mass/volume of the sample greatly influences the reported concentration for a sample, especially when contaminants are heterogeneously distributed throughout the parent matrix.

• It is like putting a drop of dye in water in a water glass vs. in water in gallon jug. The water in the glass will have a more intense color than the water in the jug.

• For heterogeneous samples (which are affected by the nugget effect to a greater or lesser degree), the analytical result for a sample is determined by how much contaminant (in the form of concentrated nuggets) is captured in that sample amidst a volume of cleaner matrix.

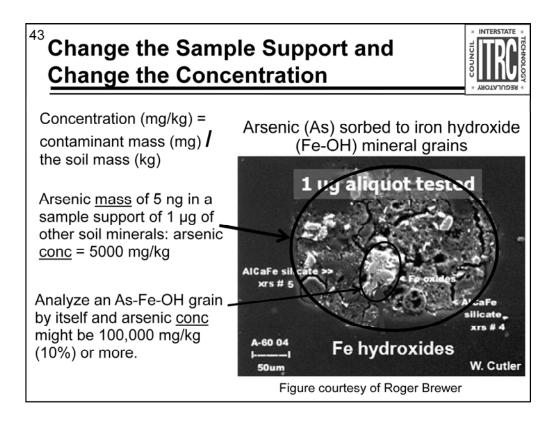
• The cleaner matrix serves to "dilute" the concentrated particles during sample extraction (for organics) or digestion (for metals).

• The issue of sample support is becoming an increasingly important determinant of analytical results as more sophisticated analytical technologies and efforts to reduce generation of lab waste drives a trend toward smaller and smaller masses of sample.



• Another way sample support influences concentration results is whether the sample support is large enough to accurately capture the particle ratio of the population.

- In this cartoon, the large container represents a field sample in a jar.
- The cartoon illustrates how subsample support affects how well a lab subsample represents the field sample.
- The lab subsample represents the field sample if the ratio of contaminant-laden particles to "cleaner" particles in the subsample mirrors the ratio in the field sample.
- A sampling error occurs when a subsample does not have same ratio as the field sample.
- Sampling error is more likely for smaller subsample supports, since they are more likely to under- or over-estimate the proportion of "hot nuggets" to less contaminated "cool" particles.
- Larger supports are more likely to represent the actual ratio and give a concentration result that is representative of the mean of the jarred field sample.
- Figure adapted from EPA 530-D-02-002, RCRA Waste Sampling Draft Technical Guidance, August 2002, page 92.
- ISM addresses this problem by collecting many increments which results in a large mass representing the whole volume of the material being investigated.



• Nuggets carrying high contaminant loading have a huge effect on what the concentration is reported to be. Concentration is determined by 2 things: the mass of the contaminant and the mass of the material that contains the contaminant.

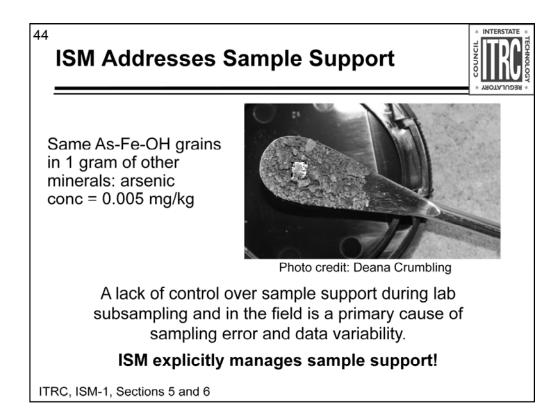
• A smaller mass of soil that contains some contaminant-laden nuggets will have a higher concentration than if the same nuggets are present in a larger mass of soil.

• In the picture, the mass of arsenic on the Fe-OH mineral grains was measured to be 5 nanograms. The mass of the soil minerals containing the arsenic is 1 microgram (blue circle). Expressed in common concentration units, 5 ng arsenic in 1 microgram of soil material is 5000 mg/kg.

• On the other hand, consider if only the arsenic-coated iron hydroxide particle itself were analyzed (red oval). The arsenic might make up 10% of the mass, while iron hydroxide makes up the rest (90%). Then the arsenic concentration would be 100,000 mg arsenic/kg of soil material.

• When the number of contaminated particles (i.e., the mass of contaminant) stays the same, the concentration will be different depending on how much soil material is digested and analyzed.

• The notion of "maximum concentration" is meaningless unless a sample support is specified. This applies in the field as well as in the laboratory.

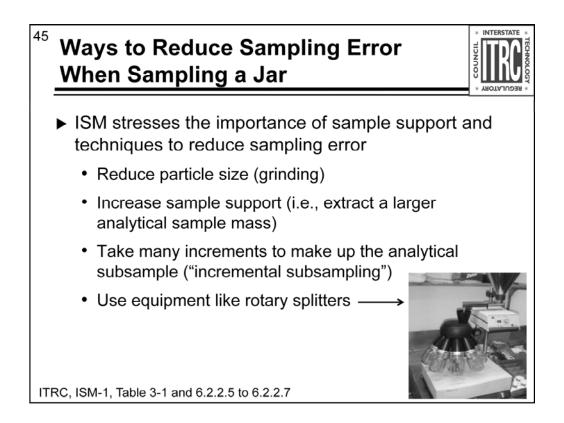


• But what if those same arsenic-bearing grains were present in 1 gram of "cleaner" soil particles (particles that are not laden with arsenic)?

- Then the arsenic concentration would be 5 ng arsenic in 1 gram of soil or 0.005 mg/kg.
- A key concern of ISM is controlling sample support!

Supplemental Information

See ISM-1 Chapters 5 and 6



• Unlike routine discrete sampling programs, ISM specifically addresses sample support issues. A project team using ISM <u>must</u> consider the likelihood of nuggets, the analytical subsample's volume and particle size.

• Reducing the overall particle size by grinding prior to subsampling may sometimes be required.

• Increasing the mass of the subsample and incremental subsampling are common ways to reduce subsampling error.

• If a field sample needs to be split, there are specialized equipment and techniques, such as rotary splitters. Choice of technique is heavily dependent on soil properties.

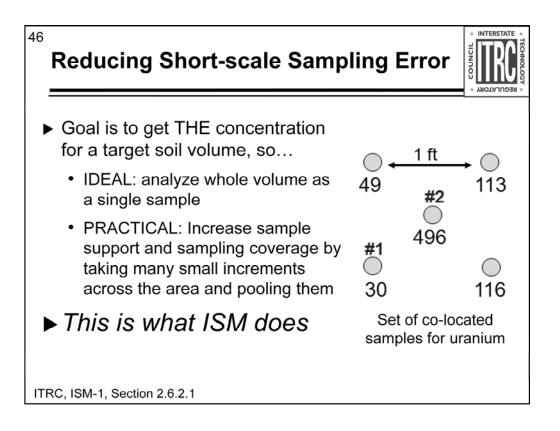
Supplemental Information

See ISM-1 Chapter 6

See also EPA guidance documents:

• "Guidance for Obtaining Representative Laboratory Analytical Subsamples from Particulate Laboratory Samples", EPA/600/R-03/027 (Nov 2003); and

• "RCRA Waste Sampling Draft Technical Guidance", EPA 530-D-02-002 (August 2002), Chapter 6



The same principles apply to short-scale sampling error. Recall that this refers to extrapolating a single data point to a large field area without taking spatial heterogeneity into account.

Taking the whole targeted soil volume as a single sample for analysis would provide THE concentration for that volume without any sampling error. But, of course, that is not possible, so we take samples.

Need to have enough samples to include fluctuations in concentration in the result for the soil volume, but without exorbitant cost.

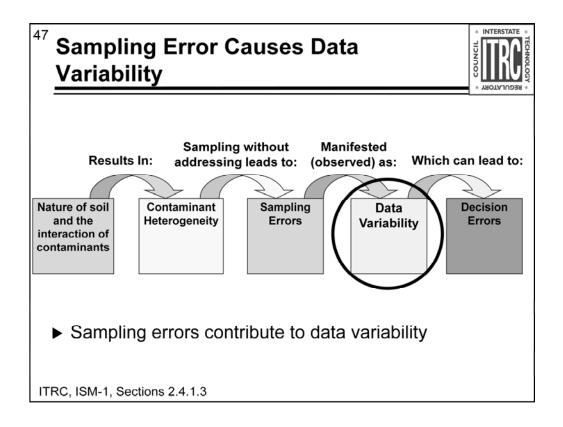
This goal can be accomplished by taking increments from many locations and pooling them together for a single analysis.

Incremental field sampling increases the sampling density (the number of samples per unit area) AND it increases the sample support of the field sample—both of which help control sampling error.

This is what ISM does in its planning stage and field implementation stage.

Supplemental Information

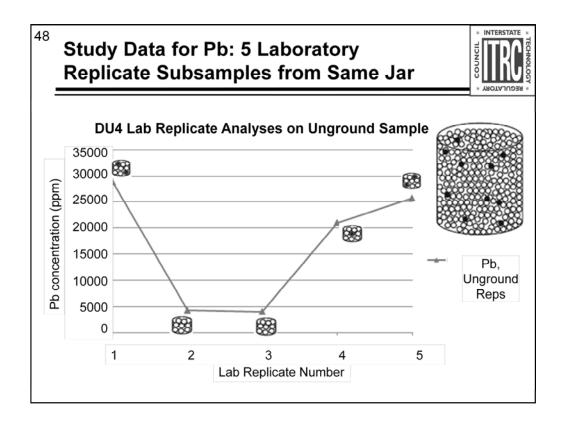
See ISM-1 Section 2.6.2.1



• Using our navigation sequence, we see that sampling errors are commonly observed as data variability.

Supplemental Information

See ISM-1 Sections 2.4.1.3



• Don't dismiss this data just because all the concentrations are high. Just because these Pb concentrations are much greater than the common risk-related threshold of 400 ppm does not mean that variability at these high concentrations are not important. Decisions about remedy selection and design or soil treatment and disposal may still hinge on differences at these high concentrations.

• The prime purpose of this graph is to illustrate the extreme variability that soil contamination can display.

• Soils that are contaminated are more likely to display a nugget effect which manifests as high variability.

• Soils that are not contaminated (or very lightly contaminated) are less likely to have particles with high contaminant loading, and so typically show less variability.

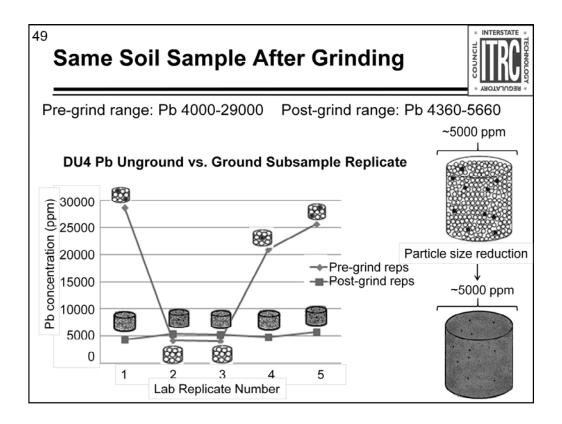
• Data variability is striking in this experiment where 5 replicate subsamples were taken from a single unground sample. Each of the 5 subsamples were analyzed for metals. The mass of the subsamples was 2.5 grams.

• The Pb results varied between 4000 and 28,000. Remember! These are not different field samples...they are 5 different subsamples from the same jar of soil.

• Fortunately, routine lab quality control checks provide measures of variability. QC includes co-located samples, field splits, lab duplicates, and matrix spike/matrix spike duplicates.

• Unfortunately, the information provided by these QC results is greatly under-appreciated and often ignored.

• A small sample mass composed of large particles frequently does not preserve the proportion of constituents as is present in the original population.



• This is a continuation of the previous slide, now with ground results for the same sample ("post-grind", in pink) also included in the graph.

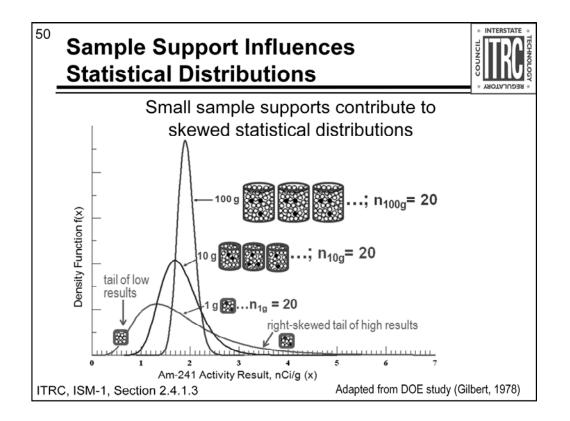
• 5 replicate subsamples were taken for analysis after the sample had been ground. The mass of the subsamples was again 2.5 grams.

• Variability was markedly reduced, which is the same as saying precision was markedly increased.

• The dramatic influence of nugget effects in the unground sample is evident by comparing the 2 sets of replicates.

• This data illustrates how grinding provides the smaller particles and mixing needed to better preserve the sample's constituent proportions even when small subsamples are used.

• The larger the particle size in the sample, the more subsample mass is needed to produce a representative subsample.



• This graph plots the data from a study done in the 1970s. It directly measured how different masses of analytical samples (i.e., the sample support) influenced the statistical distribution of the data.

• Measurement units are in nCi/g, a measure of radioactive activity, which is related to the concentration (in ppm) of the radioisotope, in this case americium-241.

• The experiment involved first preparing a large soil sample (with mass of several kilograms—not shown on the slide—which will be called a batch) from which subsamples of various sizes could be taken (as shown on the slide). Preparing the large batch involved moderate homogenization efforts involving mild grinding and then sieving to less than 10-mesh.

• A series of 20 subsamples each of different supports were taken from the large prepared batch.

• The subsample supports that were tested included 1-gram, 10-gram, and 100-g ram.

•The wider the peak shape, the more variability present in the data set.

• The data set from the 1-g subsamples plots as a statistical distribution that is unsymmetrical and skewed in that the right-hand tail is pulled out.

• The 1-g tail does not reach the x-axis until about 5 (note the green subsample on the right with a higher nugget:matrix ratio than the ratio in the 100-g samples).

• Many samples have low concentrations, reaching down to about 0.25 (green subsample on the left without any high-load nuggets)

• The width and shape (a low hump) of the curve mean that repeated 1-gram subsamplings of the large batch will produce data results that have a wide spread in values. Frequently there are low results, but sometimes there will be very high results. This variability is also called imprecision. No single result can be trusted to be close to the true mean of the batch.

• In contrast to the 1-g subsamples, the 20 10-g subsamples (purple) showed much less skewing of the right tail.

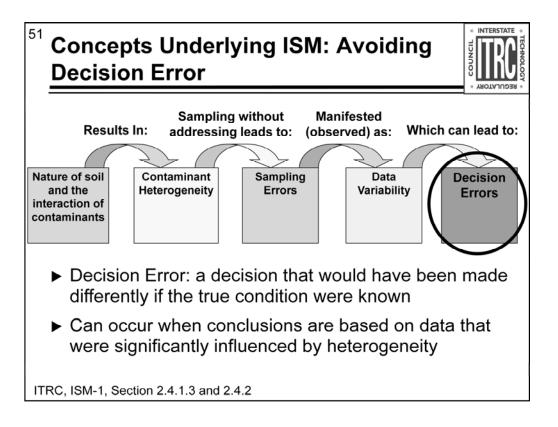
- . The right-hand tail reaches the x-axis just past 3.
- The left-hand tail shows fewer samples (than the 1-g data set) with very low results, with the lower range of the distribution ending at about 0.8
 The width of the 10-g peak is narrower, reflecting less variability (more precision) in the 10-g data set
- For the 100-g subsamples (red), the statistical distribution is almost symmetrical, with a high tight peak (high precision) and the right skewing nearly gone.
 - The 100-g curve reaches the x-axis on the right at about 2.5
 - On the left, the 100-g curve runs only down to about 1.4
 - The height and narrowness of the 100-g peak indicates that replicate subsamplings of the batch produce values that are close to each other (precise), and most likely close to the true mean for the large batch.

• Not only do small sample supports increase variability, they also contribute to data taking a lognormal or gamma (or other skewed) statistical distribution.

• So what does this have to do with decision errors?

P.G. Doctor and R.O. Gilbert. 1978. DOE NAEG Report. Two Studies in Variability for Soil Concentrations: with Aliquot Size and with Distance [provided in webinar References]

See also Gilbert, Richard O. and Pamela G. Doctor. 1985. Determining the Number and Size of Soil Aliquots for Assessing Particulate Contaminant Concentrations. Journal of Environmental Quality Vol 14, No 2, pp. 286-292.

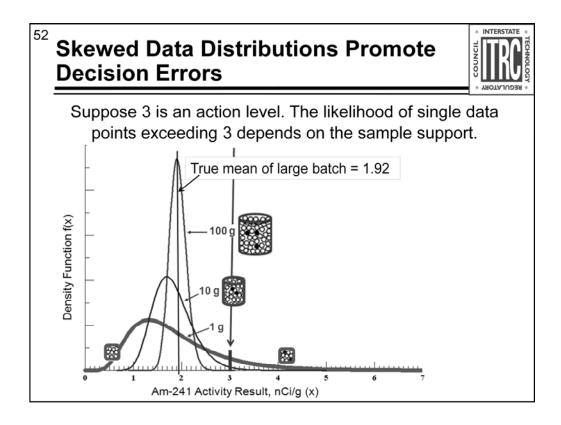


• A decision error is a conclusion that is different from the conclusion the data user WOULD have made if the true condition were known.

• Decision error in this context refers to using data results to draw a conclusion without taking data variability and other sources of sampling and analytical error into account.

Supplemental Information

See ISM-1 Sections 2.4.1.3 and 2.4.2



• It is known that the true "concentration" of the large multi-kg batch is 1.92 (refer back to a previous slide)

• Measurement units are in nCi/g, a measure of radioactive activity, which is related to the concentration (in ppm) of the radioisotope, in this case americium-241.

• For the sake of discussion, suppose 3 is an action level, which is here shown as the small vertical blue line on the x-axis.

• Therefore, the true "concentration" of the large batch (1.92) is below the action level of 3

• The question for a data user is: Will the subsample that is analyzed lead to the correct conclusion about whether the "concentration" of the batch is higher or lower than 3, or could the data lead the user astray?

• Look again at the curve representing the 1-g subsamples (the heavy-lined curve): Even though the true mean is well below 3, the skewed nature of the data means that sometimes (around 11% of the time) data results are going to be higher than 3, as exemplified by the green subsample on the right. This would contribute to a decision error.

• Note that there is a 12% chance that a 1-g subsample would have concentrations much lower (less than 1 nCi/g) than the true mean, as exemplified by the green subsample on the left.

• Look at the curve representing the 10-g subsamples (the purple subsample): Only rarely will a result from a 10-g subsample exceed 3.

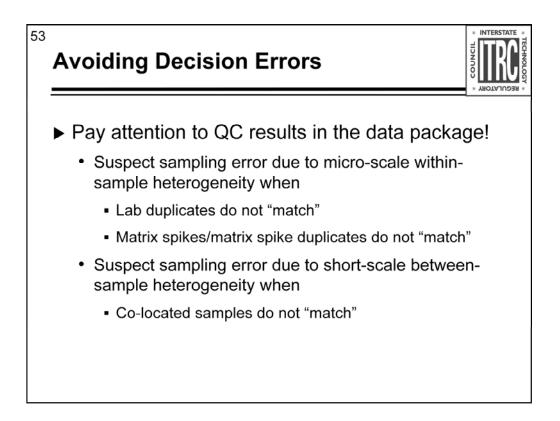
• In contrast, look at the 100-g curve (red subsample). Since that curve ends around 2.5, it is very, very unlikely that any single data result would be greater than 3.

• Larger subsamples are more likely to provide data results that are close to the true mean, as evidenced by the tighter peaks the 10- and 100-g subsamples show around the true mean.

• The bottom line is that decisions that are based on a single sample result are more likely to be in error when subsample supports are small.

• As we talked about before, metals analysis typically uses around 1 gram of soil. Deciding that a few high results represent hotspots could well be decision errors due to the skewed distribution of data from small subsamples. This is why areas initially called hotspots sometimes cannot be found upon repeat sampling.

• Sampling errors operate in the other direction too. A sample from a true hotspot might give a data result biased far lower than the true value (as illustrated by the "clean" green subsample on the left) and the hotspot would be missed.



• How can we avoid decision errors?

• When laboratory duplicates and/or matrix spike/matrix spike duplicates do not match and there is wide variation in results within the data set, suspect that sampling error may be occurring at the within-sample level.

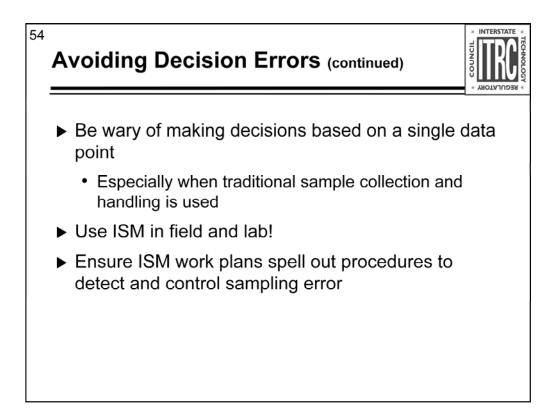
• If within-sample heterogeneity has been controlled, but co-located samples do not match, the problem is likely short-scale heterogeneity.

• When sampling error has affected a data set, making decisions based on single sample results is like flipping a coin—it is a matter of chance.

• Decisions need to be based on the data set as a whole. If the data set is large and decisions are based on the mean, or the UCL on the mean, at least some of these errors could cancel out. But typical discrete data sets are much too small for that to happen.

• So work plans such as Quality Assurance Project Plans (QAPPs) need to be constructed with procedures that control for heterogeneity's effects and measure the degree of sampling error present.

• According to EPA Superfund guidance, data error must be measured for data to be definitive. (USEPA *Applicability of Superfund Data Categories to the Removal Program* OSWER 9360.4-21FS EPA 540-F-05-005 July 2006.) So QAPP reviewers should recommend that sampling error be quantified and controlled.



• How can we avoid decision errors?

• When laboratory duplicates and/or matrix spike/matrix spike duplicates do not match and there is wide variation in results within the data set, suspect that sampling error may be occurring at the within-sample level.

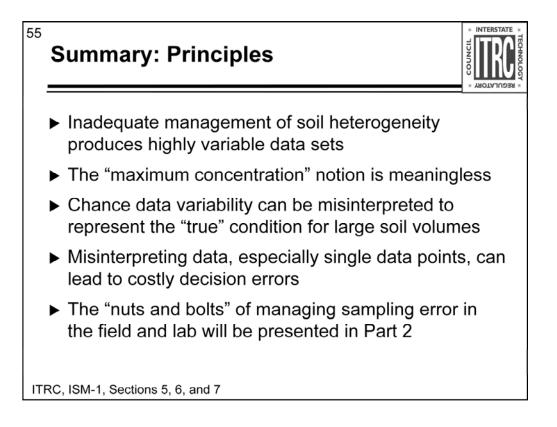
• If within-sample heterogeneity has been controlled, but co-located samples do not match, the problem is likely short-scale heterogeneity.

• When sampling error has affected a data set, making decisions based on single sample results is like flipping a coin—it is a matter of chance.

• Decisions need to be based on the data set as a whole. If the data set is large and decisions are based on the mean, or the UCL on the mean, at least some of these errors could cancel out. But typical discrete data sets are much too small for that to happen.

• So work plans such as Quality Assurance Project Plans (QAPPs) need to be constructed with procedures that control for heterogeneity's effects and measure the degree of sampling error present.

• According to EPA Superfund guidance, data error must be measured for data to be definitive. (USEPA *Applicability of Superfund Data Categories to the Removal Program* OSWER 9360.4-21FS EPA 540-F-05-005 July 2006.) So QAPP reviewers should recommend that sampling error be quantified and controlled.



• In summary: Inadequate management of soil heterogeneity produces data sets tainted by sampling errors that manifest as data variability where data consistency would be expected.

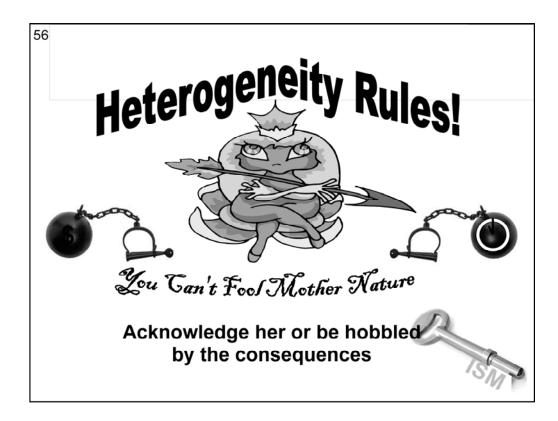
• If single data results or very small data sets are used to make decisions, data variability and chance can produce data that might be misinterpreted as the "true" condition for large volumes of soil.

• Misinterpreting data sets can lead to costly and non-protective decision errors about risk, compliance and remediation of soils.

• Controlling sampling error is a prime feature of ISM, and more information on this will be presented throughout this webinar and in the ITRC ISM Tech-Reg document.

Supplemental Information

See ISM-1 Sections 5, 6 and 7



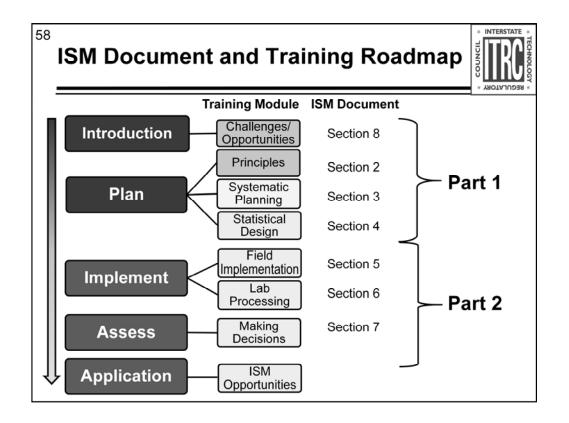
• As a bit of whimsy, this slide is meant to convey that heterogeneity is the natural state for soils.

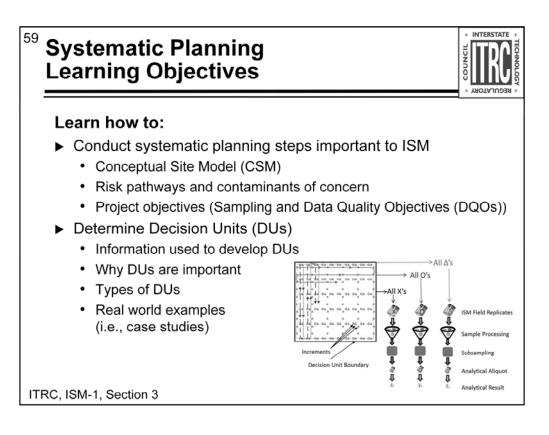
• Pretending that heterogeneity doesn't exist will hobble our projects with wasted time and money.

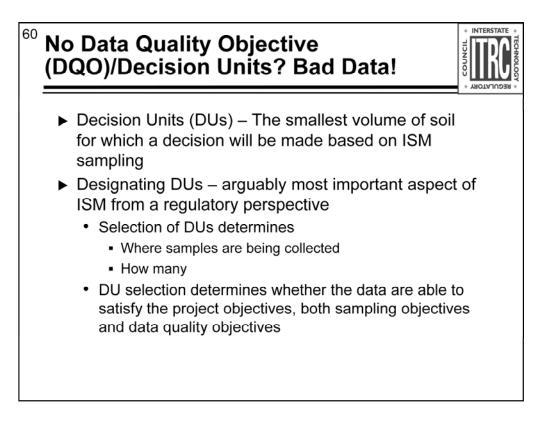
• Incremental sampling methodology is key to managing micro-scale heterogeneity to reduce subsampling error in the lab, as well as increasing sampling densities to manage short-scale heterogeneity in the field.

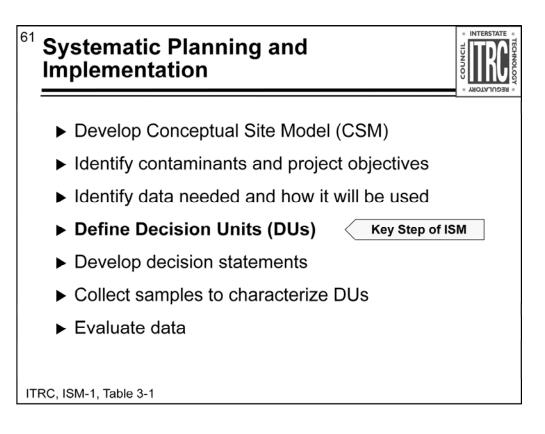
The next section of the training will delve into another aspect of ISM that must be carefully planned, which is how to select the appropriate decision unit (DU) size, configuration, and location.



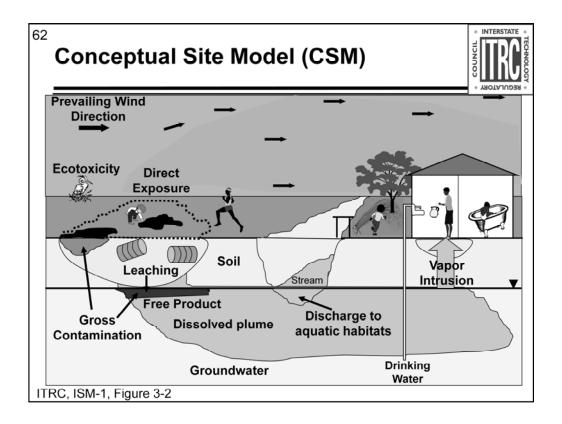






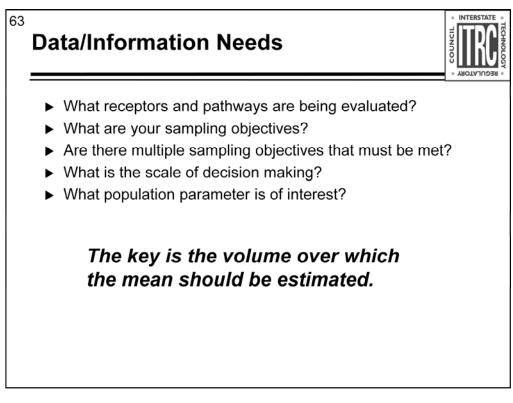


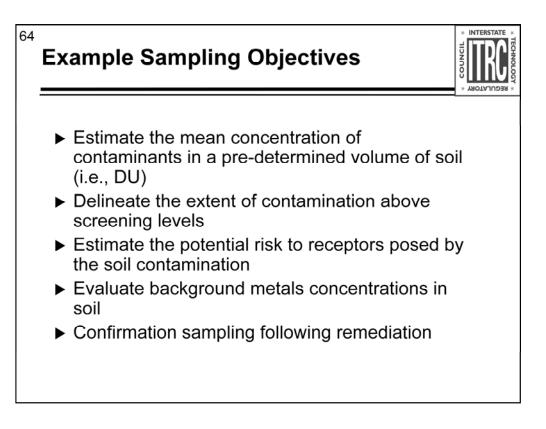
Refer to Table 3-1 in ITRC ISM-1.

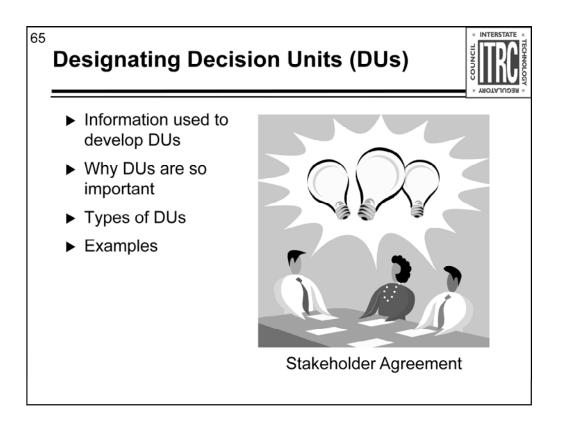


Typical Information: Historical Site Use - Current and Future Receptors – Contaminants of Concern – Identify Potential Source Areas – Evaluate Migration Pathways – Goals of the Investigation – Geologic Conditions

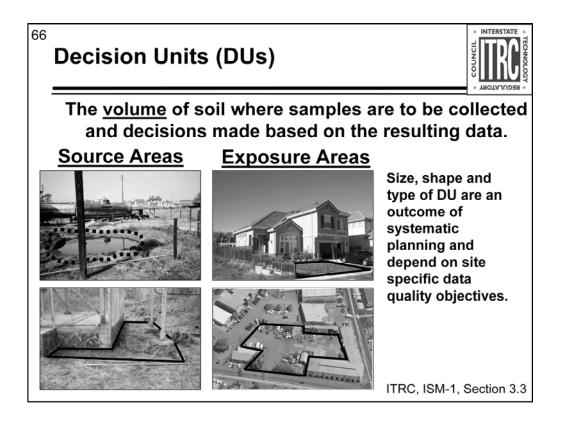
ITRC ISM-1: Figure 3-2



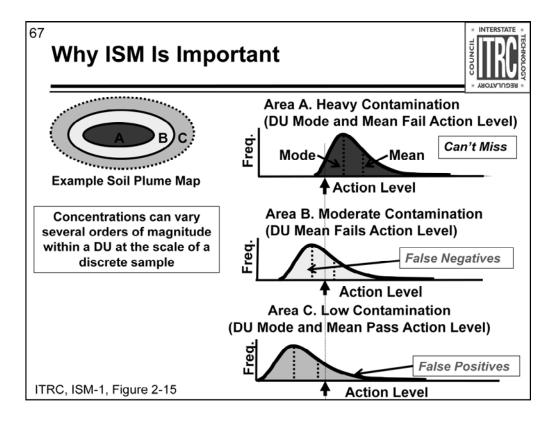




Information that can be used to determine DUs includes: Historical site use; aerial photos for possible source areas; existing sampling data; interviews with current or former site workers; sampling objectives; and data quality objectives.

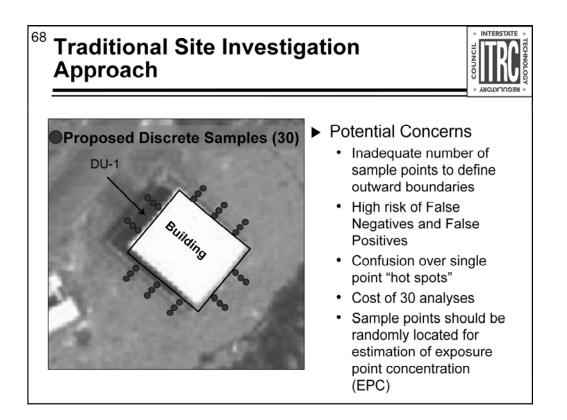


Refer to ITRC ISM-1 Section 3.3.1 Exposure Area Decision Units and ITRC ISM-1 Section 3.3.2 Source Area Decision Units

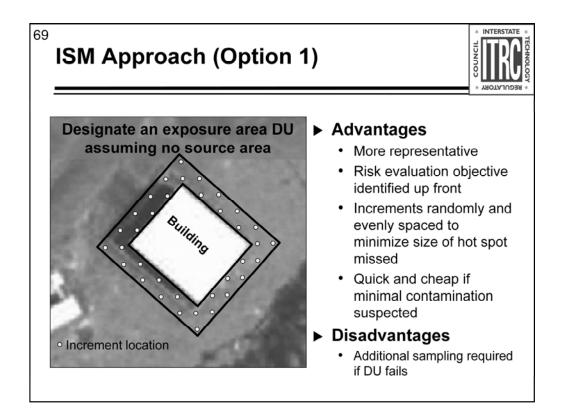


Source areas can be found with discrete data, however boundaries are much more difficult. As the mean gets closer to the action level, the greater the chances discrete data will miss contamination and underestimate the mean. Take home message: Don't use discrete samples to estimate the extent of contamination!

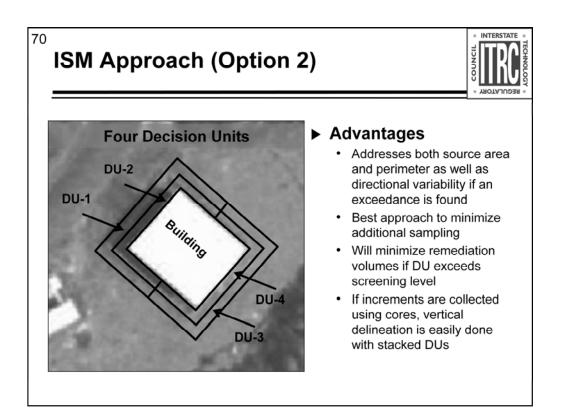
ITRC ISM-1: Figure 2-15.

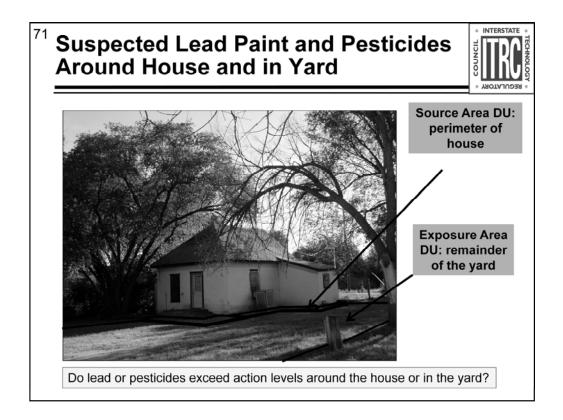


Simple discrete design. Poor at identifying site boundaries, high risk of false negatives and false positives, potential to consider a single data point a hot spot, more expensive, and not a good design for estimating risk for an exposure area.

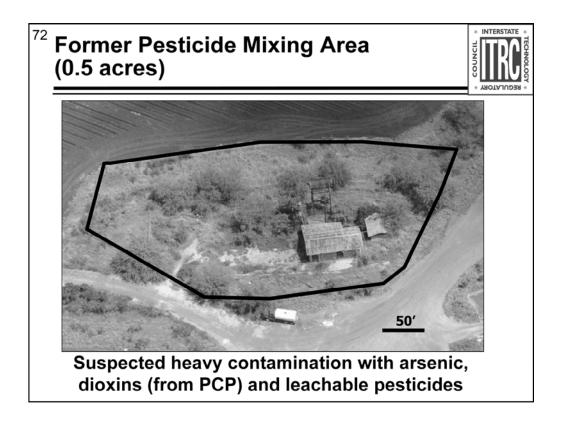


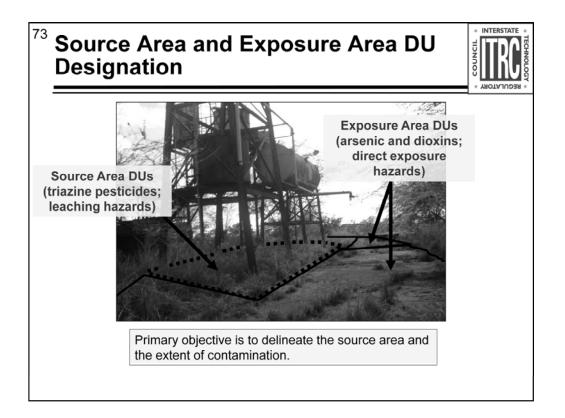
The Decision Statement: "Does contamination in soil around the perimeter of the building pose potential direct exposure concerns?" is the same as it was for the discrete design on the previous slide. This is agreed upon before the investigation. If the mean concentration is lower than a target screening level then no further action will be required. If the mean concentration will be required.



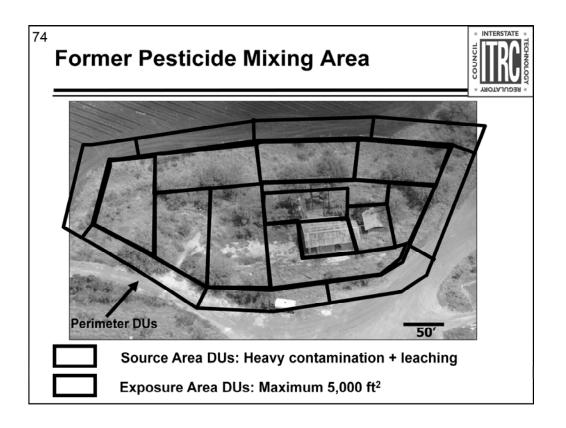


Example Decision Units for estimating the concentration of lead from lead-based paint around a home as well as pesticides in the yard.



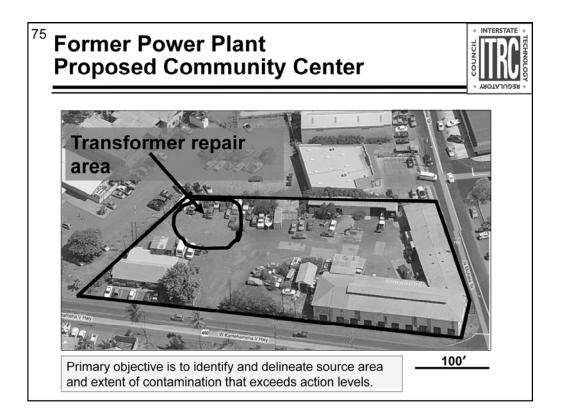


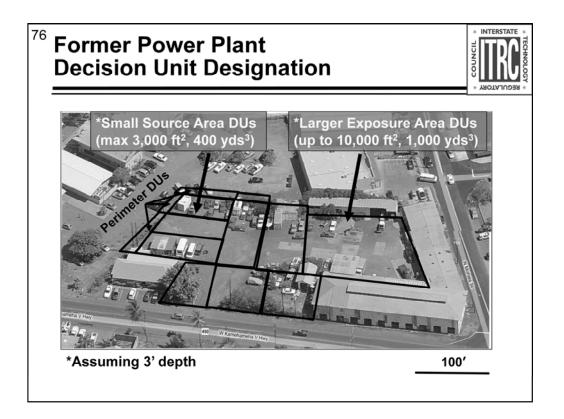
Use of small source area DUs and larger exposure area DUs for estimating the boundaries of contamination.

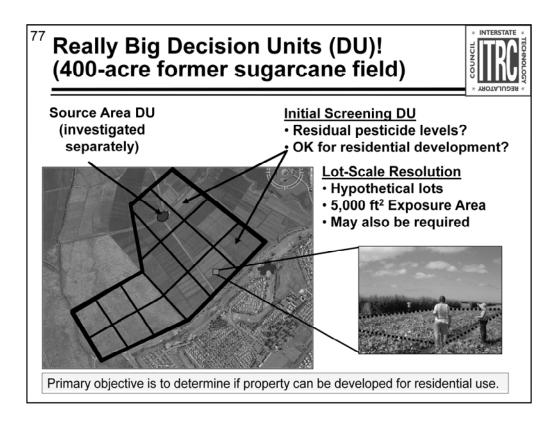


Exposure area DUs based on a default residential lot size.

This is how the scale of decision making is tied directly to the sampling design.





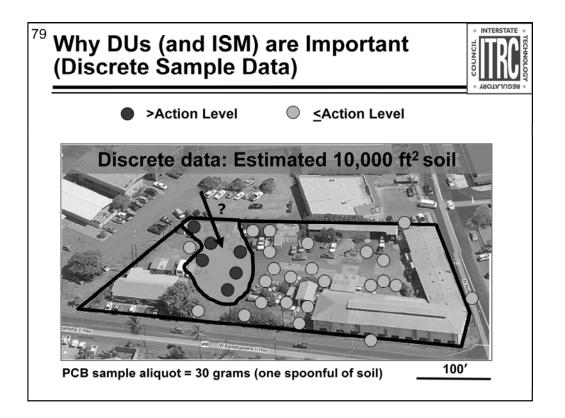


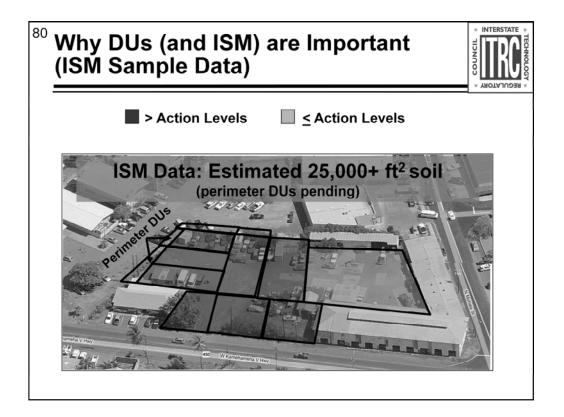
Refer to ITRC ISM-1 Section 3.3.7.

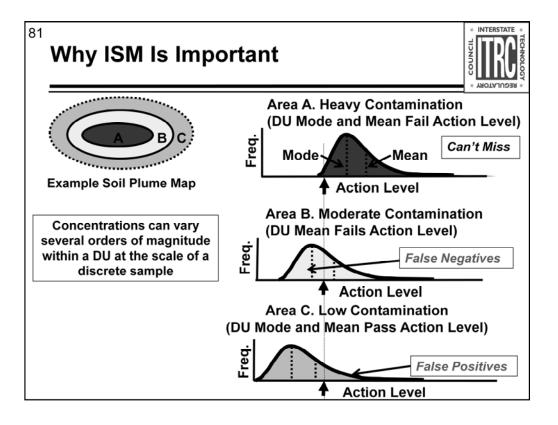
⁷⁸ Really Small Decision Units??? What about the Sandbox!?

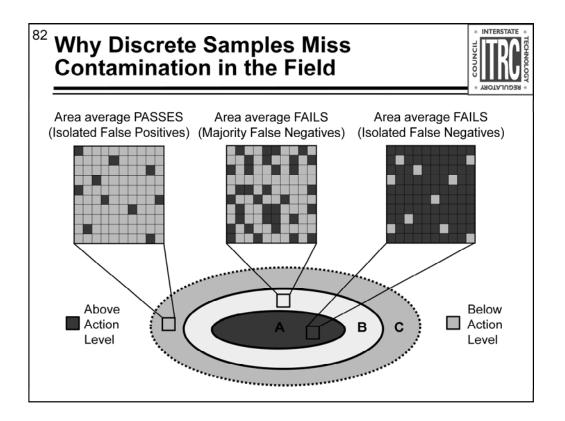


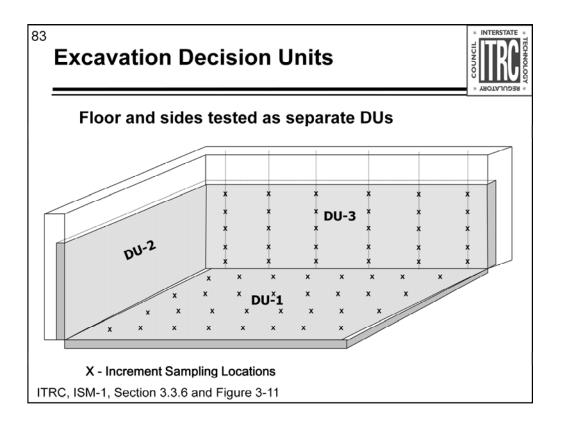
- Yard-size DUs are most often appropriate
 If acute hazards or intense exposure are being evaluated, smaller DUs may be necessary
 Not typical
 - Investigate known or suspected source areas separately
 - Remember: As sampling objectives change, so must the sampling design



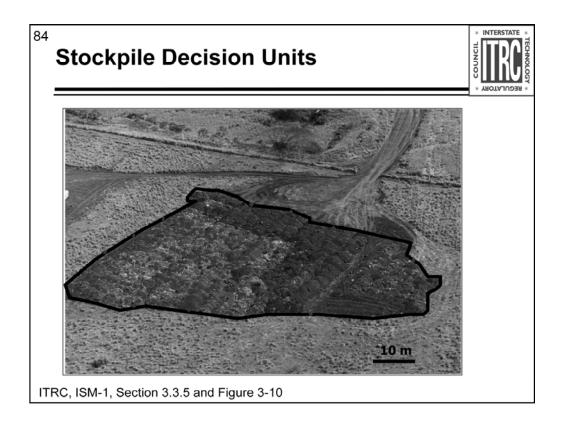






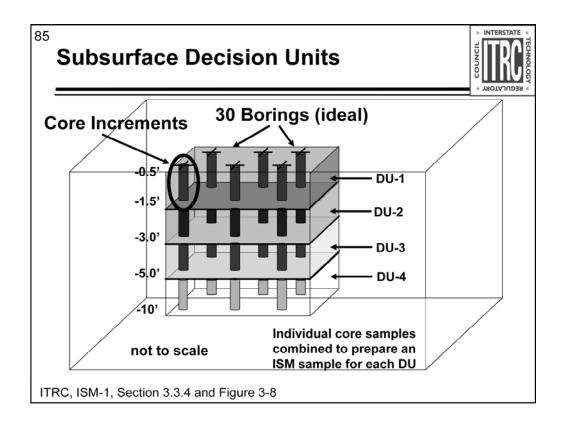


Refer to ITRC ISM-1 Section 3.3.6 and Figure 3-11

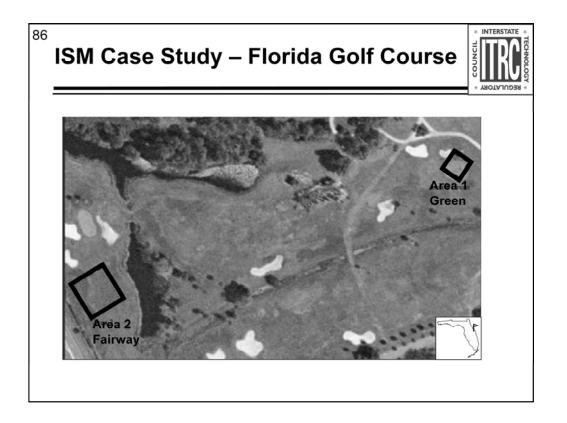


The soil from the stockpile was to be used for fill material in a residential development, and spread out to a depth of six inches over 5,000 square foot lots. This is approximately 100 cubic yards of soil per residential lot. The stockpile was then divided into 100 cubic yard DUs.

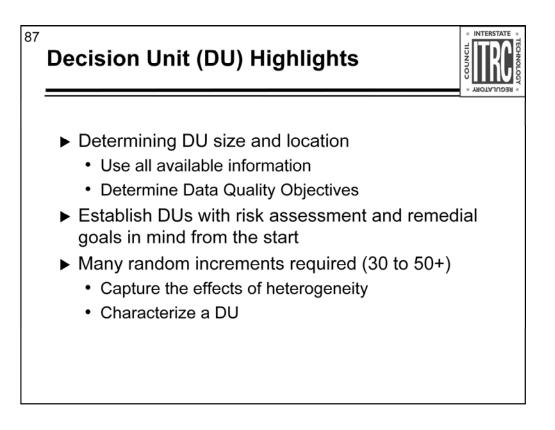
Refer to ITRC ISM-1 Section 3.3.5 and Figure 3-10

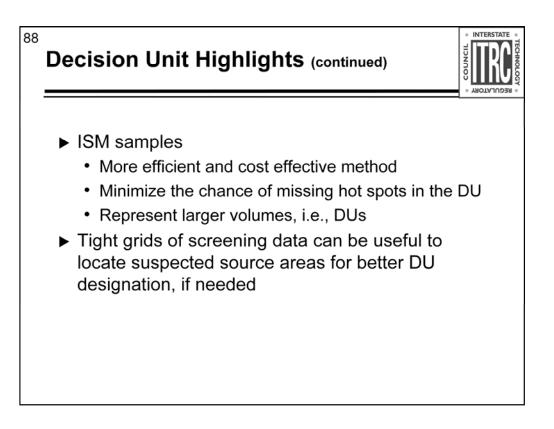


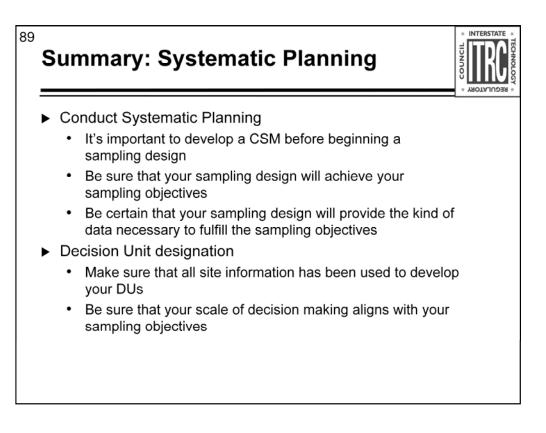
Refer to ITRC ISM-1 Section 3.3.4 and Figure 3-8

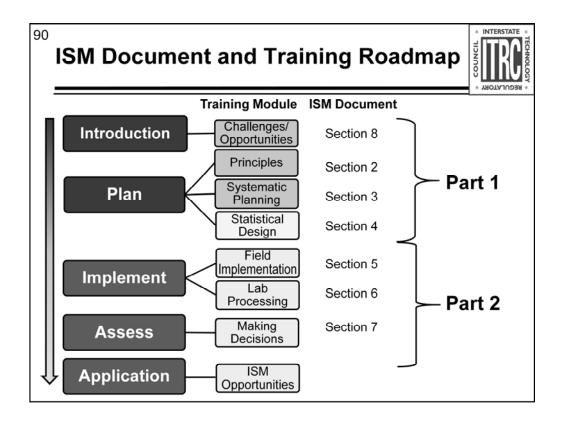


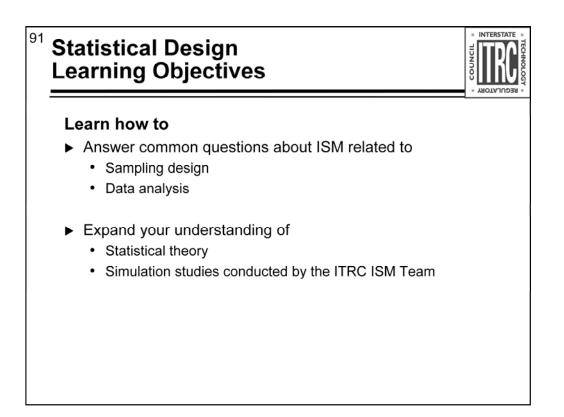
Decision Units designed to test various aspects of ISM, not characterize the entire golf course.





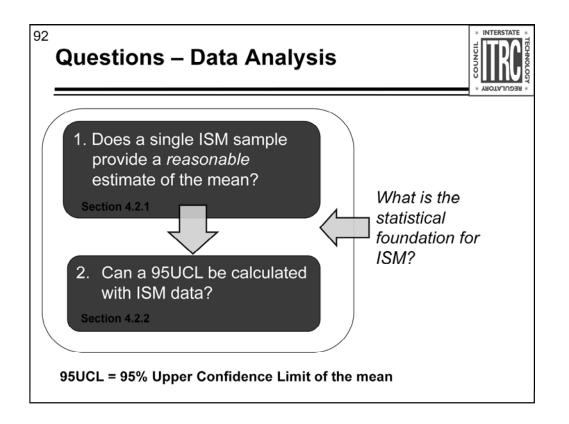






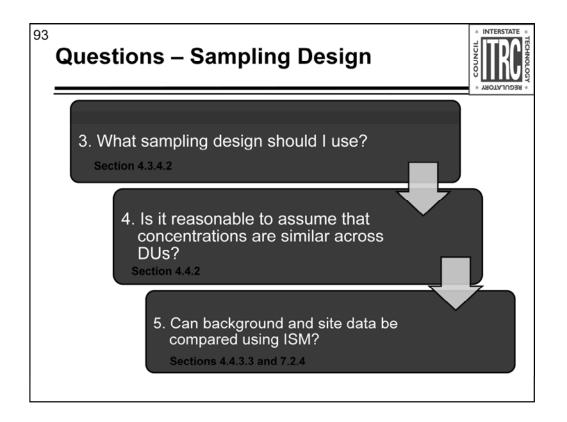
Access additional information on statistical design for ISM

ITRC ISM-1 Other training modules



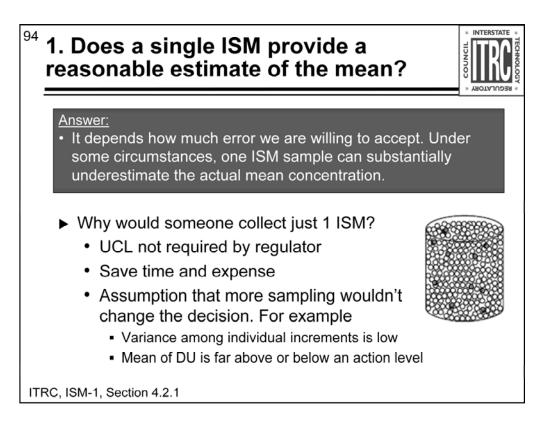
Note that section numbers of the document are given for more detailed discussions of these topics.

If the answer to #2 is yes – then the next question is - what UCL method should be used?



- Sampling designs include multiple features: sampling pattern, number of increments, and number of replicates.

- Extrapolation requires assumptions that the mean or variance are the same across multiple DUs.



Section 4.2.1

Think of each ISM result (or "replicate") as providing one point estimate from a distribution of possible means – the arithmetic mean of that distribution, also called the "grand mean", is equal to the population mean. No sampling design yields a perfect estimate of the mean. The magnitude of the error in the estimate increases as: 1) the number of replicates decreases; and 2) the variance of the distribution of means increases.

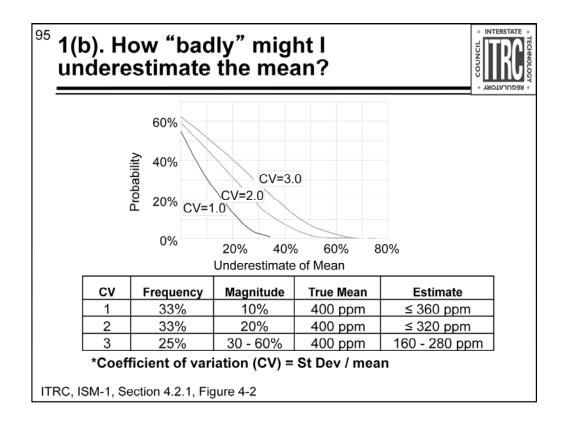
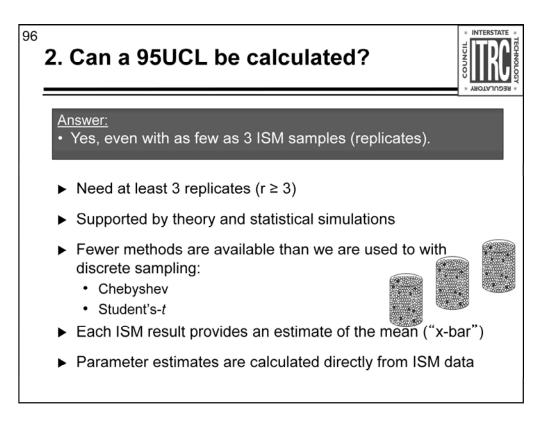


Figure 4-2, Section 4.2.1

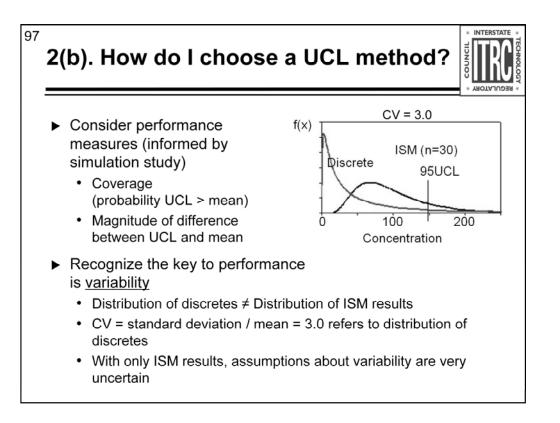
The CV in each case is based on the standard deviation of the "underlying distribution". We generally do not know this SD (and therefore, cannot calculate the CV) since we do not measure concentrations in each increment that is composited to generate the ISM. The CV may be estimated if we also had discrete sampling.



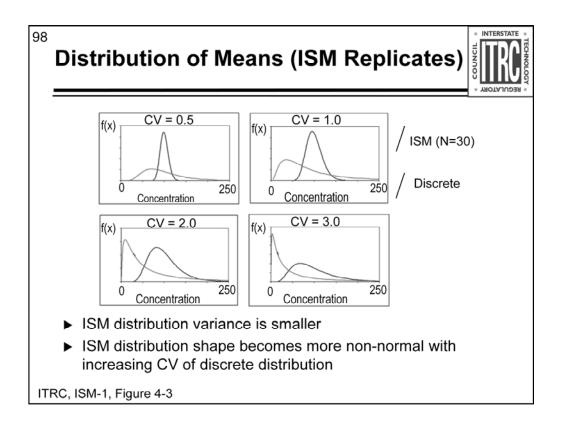
Section 4.2.2

UCL methods used with specific distributions (e.g., lognormal, gamma) would require larger sample sizes than are typically available with ISM data.

UCL methods based on bootstrap resampling would also require larger sample sizes.



In these examples, the true mean is meant to be 100. The pink distribution represents discrete data; the blue distribution represents ISM data. Chebyshev will yield a higher UCL than Student's t. This allows for better coverage but also a greater magnitude of difference between the UCL and mean. Performance varies depending on the underlying distribution's shape and variance. This makes the choice difficult because ISM data do not give much information about the underlying distribution.

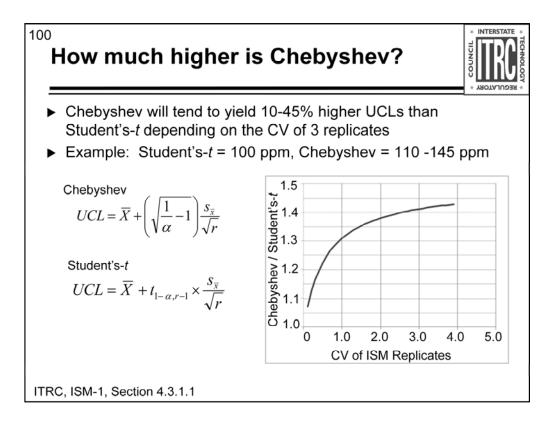


Central Limit Theorem suggests that the distribution of means approaches normality with increasing sample sizes (number of increments). However, as demonstrated here, deviations from normality are apparent for the bottom two scenarios (CV = 2 and 3). This has implications for the performance metrics of the Student' s-t UCL, which provides sufficient coverage so long as the assumption of normality (of the mean) holds true.

Coverage P	robabilit	ies	
UCL Method	Dispersion Among Individual Increments		
	Low	Medium	High
	(CV <1.5 or GSD <3)	(1.5 < CV < 3 or 3 < GSD < 4.5)	(CV >3 or GSD >4.5)
Student's-t	Yes	No	No
Chebyshev	Yes	Yes	Maybe
 Chebyshev has variability 	provide desired s more consiste	95% coverage when ent 95% coverage for r	nedium and high
C 1	, , , ,	provides marginal impl no improvement for St	
RC, ISM-1, Table 4-4,	Sections 4.3; Ap	pendix A	

This table reflects consensus results from simulation studies conducted by several members of the ISM team. This table specifically summarizes simulations of many thousands of applications of an ISM sampling protocol to hypothetical DUs. We assumed that the distribution of increments was lognormally distributed with CVs ranging from < 1.5 to > 3. ISM sampling protocols were also varied to investigate performance using different numbers of increments and replicates.

ITRC, ISM-1, Table 4-4, Sections 4.3.1.1, 4.3.3.1, 4.3.4.1; Appendix A.6.6, Fig. A-1, A-10, A-14, A-15, A-19, A-21



When both methods are applied to the same dataset, Chebyshev will yield 10-45% higher UCLs than Student's-t. For example, if the Student's t-UCL is 100 ppm, we might expect the Chebyshev UCL to be between 110ppm and 145 ppm. The exact difference depends on the variability in ISM results. Here we express that variability as the ratio of the SD to the mean (i.e., the CV). This CV of ISM replicates should not be confused with the CV of increments that was presented in the previous slide on coverage. Note that the Central Limit Theorem suggests that CV of replicates is ~ 5.5 times smaller than CV of increments / sqrt(30), and sqrt(30) = 5.48). So if CV of increments = 3.0, then CV of replicates = 0.55. At this degree of variability, Chebyshev will yield about 20% higher UCL than Student's t.





Answer:

- EPA is updating ProUCL to include an ISM module. Visit the ITRC website for additional tools.
- ProUCL was originally designed to work with discrete sample data, but is being updated to include an ISM module.
- Only Chebyshev and Student's-t UCLs are implemented for ISM datasets.
- ITRC guidance has calculator tools that work for ISM data (see ISM-1, Sections 4.2.2 at http://www.itrcweb.org/ism-1/4_2_2_UCL_Calculation_Method.html).

ITRC, ISM-1, Sections 4.2.2

ProUCL provides more options for UCL calculations, but needs n=8 to 10 observations in order to evaluate distributions or conduct bootstrap resampling.

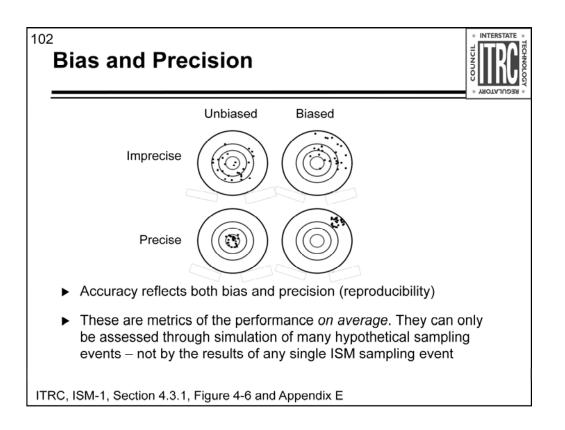


Figure 4-6 in Section 4.3.1 and Appendix E (Glossary)

Before answering this, it is important to review the meaning of bias and precision (reproducibility), and how they can both contribute to error.

• Bias = the tendency for a measurement to consistently over- or underestimate the actual (true) value. Together precision and bias determine accuracy.

• Precision (reproducibility) = a measure of reproducibility. Together precision and bias determine accuracy

Components of the RSD



Field

103

- Number of increments
- Increment collection
- Field processing
- Field splitting
- DU size and shape

Laboratory

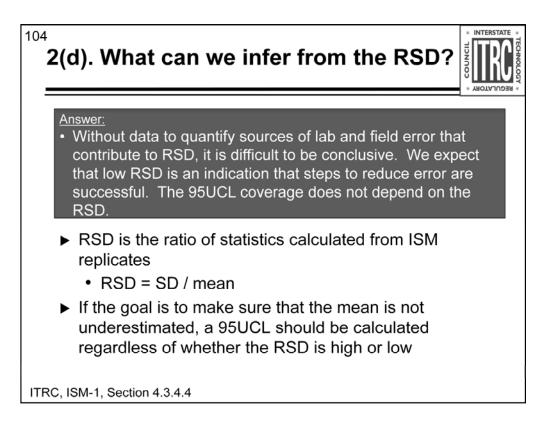
- Lab processing
- Subsampling
- Extraction
- Digestion
- Analysis
- Simulations used to explore alternative sampling designs did not attempt to isolate sources of error
- In Day 2 of ISM training, methods will be presented for quantifying and reducing relative errors associated with field and lab practices that contribute to RSD

Error (Variability) - not "mistake"

Speaker Bullets – not comprehensive; theoretical sources of error discussed in Chapter 2 (Section 2.2) ISM controls these better than discrete

It is useful when evaluating data to consider the steps in the process where errors (variability) may have been introduced

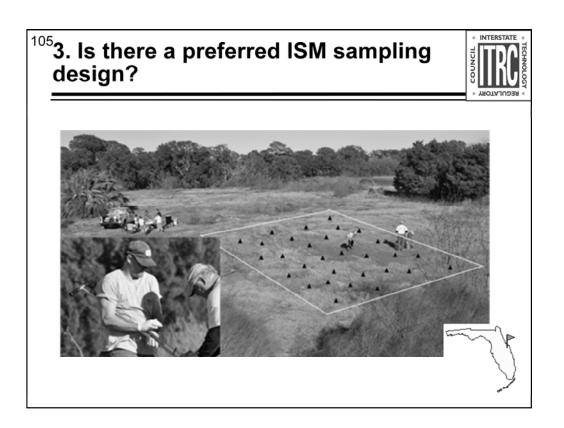
If error is determined to be unacceptable in a given data set – one or more of these sources may be at fault



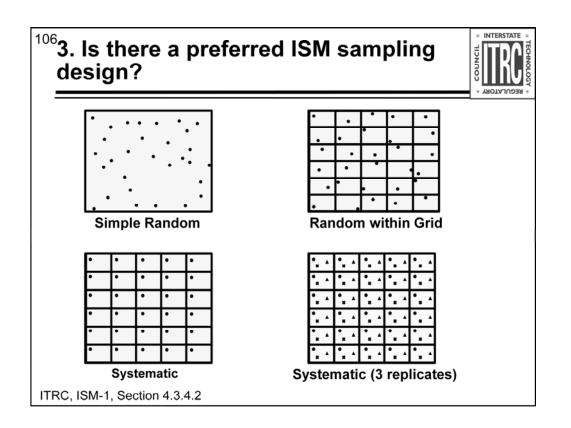
Section 4.3.4.4

RSD measures reproducibility, not accuracy of the results.

A high RSD can be caused by lab error and should be investigated. A low RSD suggests the absence of lab error, but does not necessarily indicate that the results are sufficiently accurate to avoid decision error.

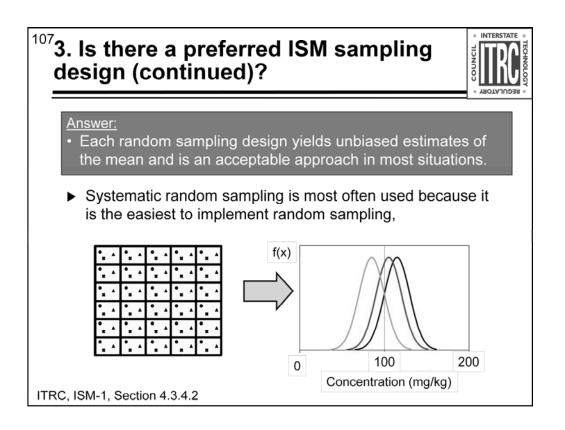


Implementation of sampling designs requires coordination between the statistician and field collection team. If the statistician provides a set of coordinates (selected at random), the field team will first place flags to match those locations in the field.



Section 4.3.4.2

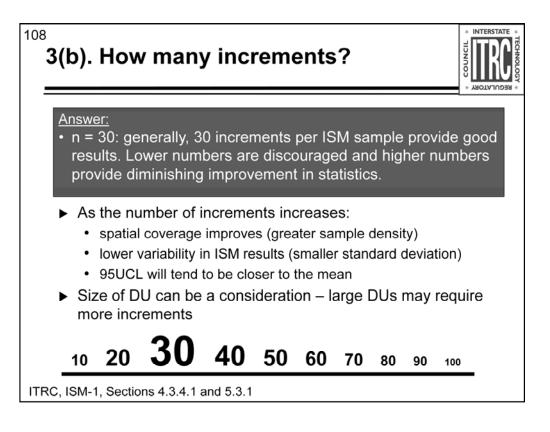
With ISM, we can consider each of the 30 points to represent an individual increment. Collectively, the set of increments are combined to form a single composite sample that yields 1 ISM result. In the four sampling designs shown, each of these patterns is a form of random sampling. This means that the exact locations of the individual increments can change with each new sampling event. With #1, even though the increments are equidistant, the location of the first increment is selected a random from within the grid cell. With #2, three sampling events are shown (circle, square, triangle), each with a different random "starting" location. It is clear how the density of the samples (or "spatial coverage") can be very high with ISM designs that involve multiple replicates.



Section 4.3.4.2 (sampling design) and Section 4.3.1.2 (bias)

Random sampling from a population generates an estimate of the mean concentration of that population with desirable statistical properties.

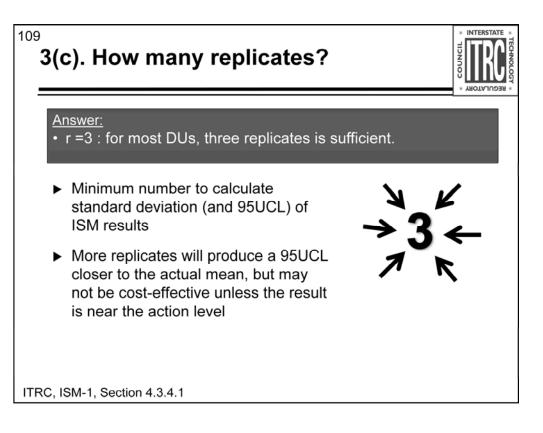
Bias is one metric used to evaluate the performance of a parameter estimation method. A sampling method is unbiased if, when repeated many times, the parameter estimate equals the population parameter. This examples shows the distribution of increments for three sampling events applied to a DU with a true (population) mean of 100 mg/kg. For purposes of presentation, we also assume that the distribution is normal. While no individual ISM is centered on the mean, on average (if repeated many times), we would expect the estimate of the mean to equal 100 mg/kg.



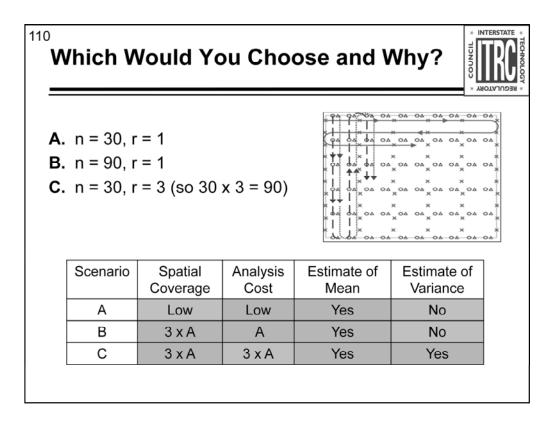
Section 4.3.4.1

Sample support and spatial coverage are usually optimized at about 30 increments. Lesser numbers compromise the ability of ISM to address sampling error; larger numbers provide limited improvement in error reduction, though a "denser" sampling grid. In the simulations, we calculated the difference between the UCL and mean using the "relative percent difference", RPD = (UCL – mean) / mean

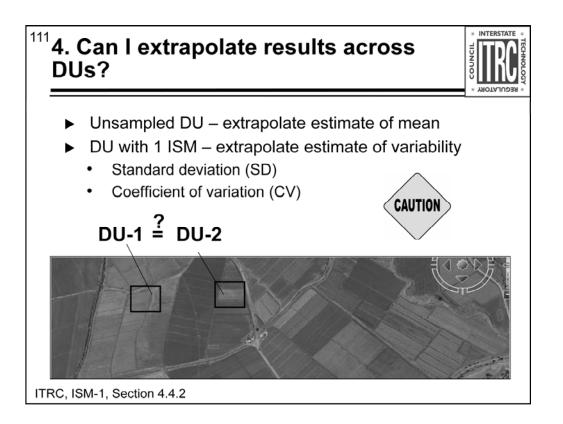
The simulations that support recommendations given in the ISM guidance apply to DUs of any size because heterogeneity was represented by statistical distributions that did not depend on the size of the DU. Some practitioners of ISM have found that increasing the number of increments to more than n=30 for larger DUs (e.g., 1 acre or more) can provide greater confidence that, collectively, the set of increments collected in the DU will represent subareas of high and low concentrations in the appropriate proportions. As discussed in Section 5.3.1 of the ISM guidance, as the DU gets significantly larger, the amount of distributional heterogeneity may increase. In these cases, depending on site specific knowledge, the conceptual site model (CSM), and data quality objectives (DQOs), it may be necessary to increase the number of increments in each DU typically reduces the variation among replicate samples. Alternatively, splitting larger DUs into two or more smaller DUs should be considered. It is not normally necessary to increase the number of increments unless there is reason to believe the DU has more distributional heterogeneity than can be controlled with 30–50 increments.



No associated notes.



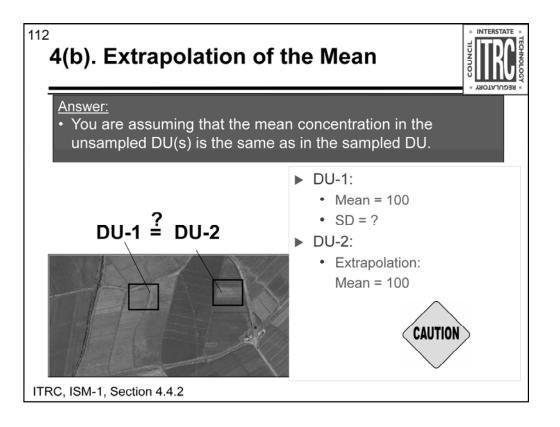
With ISM, selecting the sample sizes requires weighing advantages and disadvantages of factors such as: spatial coverage of the DU, costs (including field and analytical), and whether or not an estimate of the variance can be obtained to calculate a 95UCL. The table above compares and contrasts three scenarios. Green shading is more favorable, orange shading is less favorable. In general, we would opt for a sampling design that provides an estimate of the variance.





There are different assumptions associated with each, and therefore different answers...

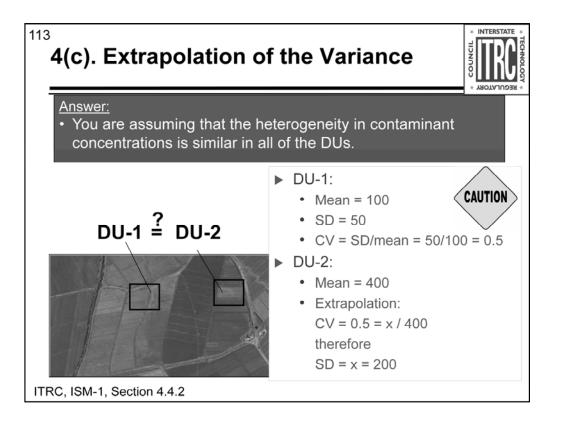
If you extrapolate variability, generally use the CV instead of the SD. This is because the CV estimates standard deviation based on the estimate of the mean: CV = SD/mean. This is more consistent with contamination that has a non-normal (or "positively skewed") distribution (e.g., lognormal, gamma). Extrapolation of the SD is only recommended if the distribution of the concentrations of individual increments is approximately normal.



Extrapolation is usually reserved for very large sites where sampling all DUs is prohibitively expensive.

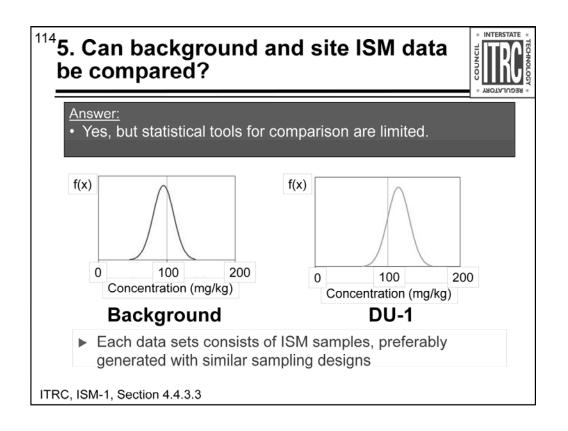
There is usually no confirmation of this assumption, so the rationale for the assumption must be very strong.

Note that there is nothing magic about ISM that diminishes the uncertainty of extrapolating from sampled to unsampled areas. If you wouldn't do it with discrete data, you shouldn't do it with ISM.



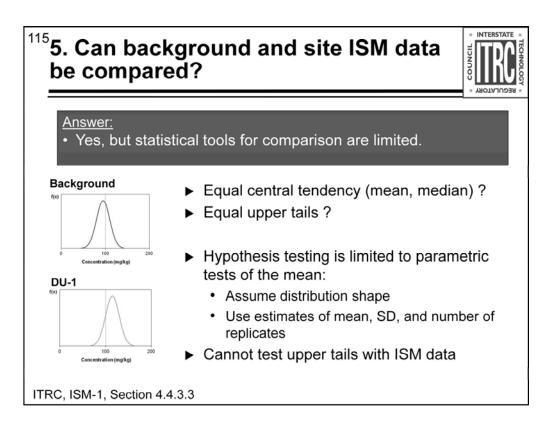
Extrapolation of variance is valid only if the distributions of concentrations in each DU are statistically similar.

If you extrapolate variability, generally use the CV instead of the SD. This is because the CV estimates standard deviation based on the estimate of the mean: CV = SD/mean. This is more consistent with contamination that has a non-normal (or "positively skewed") distribution (e.g., lognormal, gamma). Extrapolation of the SD is only recommended if the distribution of the concentrations of individual increments is approximately normal.



Section 4.4.3.3

The concept is to compare distributions. With ISM, we will generally have too few data points (e.g., r=3) to determine distribution shapes, or to use non-parametric methods. One cannot compare discrete data to ISM data because they represent the distribution of concentrations very differently.



The concept is to compare distributions of ISM results. With ISM, we will generally have too few data points (e.g., r=3) to determine distribution shapes.

Because of the small sample size, statistical power is too low to allow for non-parametric hypothesis tests like Wilcoxon Rank Sum (Mann Whitney).

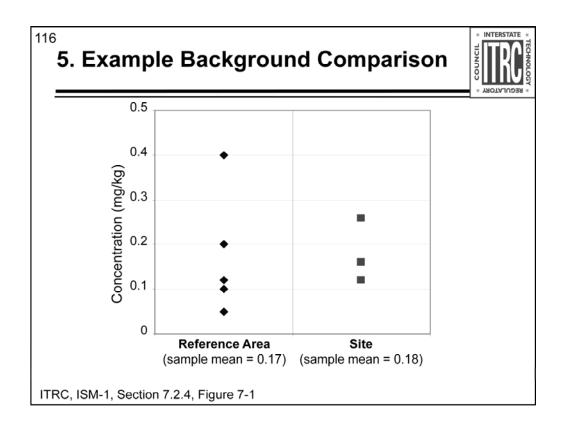
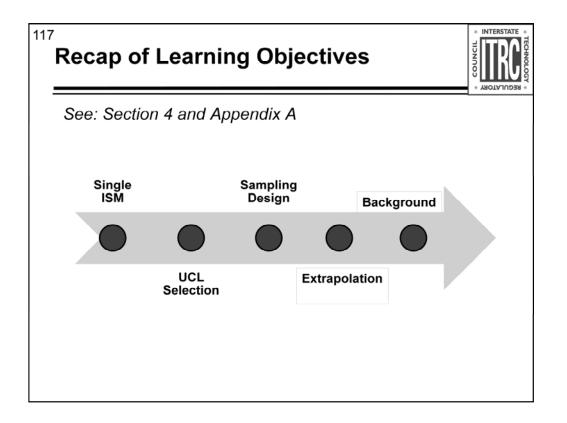
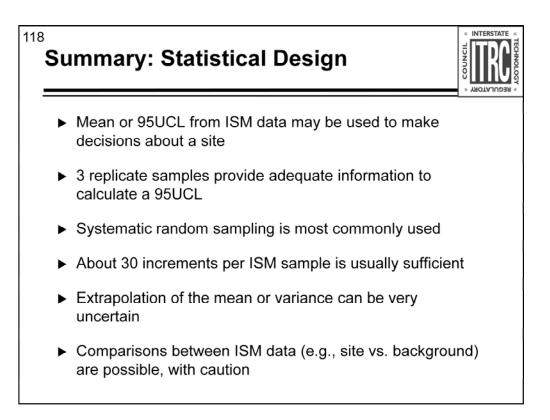


Figure 7-1, Section 7.2.4

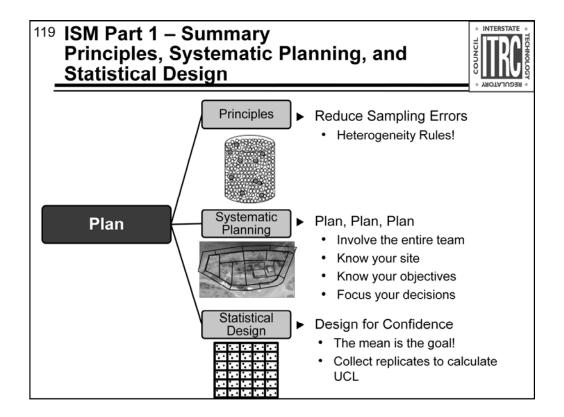
Dot plots can be a useful graphic for presenting ISM results. While not the same as a formal statistical test, the graphic presents information in a manner that can support decisions.



This Module 4 brings the Day-1 training to an end. The focus of this module has been on specific questions that can be addressed through statistical analysis of ISM data. For more detailed discussion of these concepts and an overview of the simulations that were performed to support the recommendations that have been provided, please refer to Section 4 and Appendix A of the document.



No associated notes.

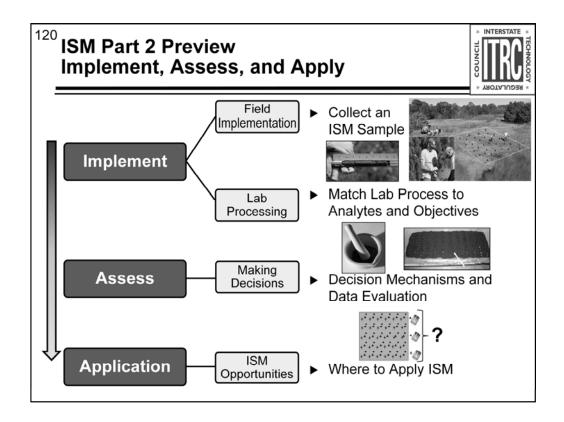


Today's training included Modules on:

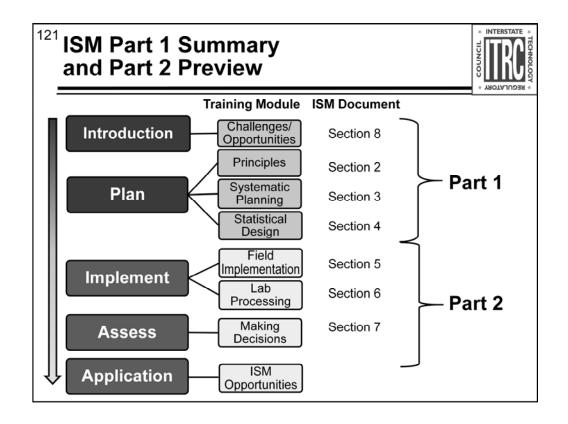
Soil and Principles - Be aware of issues related to heterogeneity and sampling errors

Systematic Planning - Involve the entire team, regulators, consultants, responsible parties in critical elements (e.g. conceptual site model, establish sampling objectives and decision units.) Sampling objectives should drive your sampling design, and the scale of decision making should align with sampling objectives.

Statistical Design - Provides the statistical foundation and describes why ISM provides a reasonable mean, describes a good ISM sampling design, and informs you how ISM provides 95% UCL.

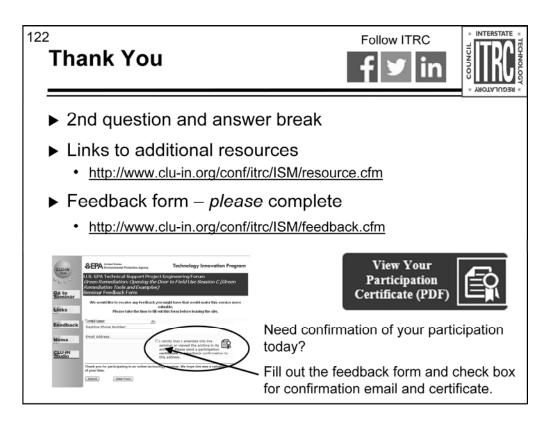


No associated notes.



Part 2 Module includes:

Collecting Field Samples Laboratory Processing & Analytical Issues Using and Applying ISM Data



Links to additional resources: http://www.clu-in.org/conf/itrc/ISM/resource.cfm

Your feedback is important – please fill out the form at: http://www.cluin.org/conf/itrc/ISM/feedback.cfm

The benefits that ITRC offers to state regulators and technology developers, vendors, and consultants include:

✓ Helping regulators build their knowledge base and raise their confidence about new environmental technologies

✓ Helping regulators save time and money when evaluating environmental technologies

 \checkmark Guiding technology developers in the collection of performance data to satisfy the requirements of multiple states

 \checkmark Helping technology vendors avoid the time and expense of conducting duplicative and costly demonstrations

✓ Providing a reliable network among members of the environmental community to focus on innovative environmental technologies

How you can get involved with ITRC:

 \checkmark Join an ITRC Team – with just 10% of your time you can have a positive impact on the regulatory process and acceptance of innovative technologies and approaches

- ✓ Sponsor ITRC's technical team and other activities
- ✓ Use ITRC products and attend training courses
- ✓ Submit proposals for new technical teams and projects