# Integrating population genomic data to understand mechanisms of chemical susceptibility and resistance

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Superfund Research Program







early life exposures, later life consequences



National Institute of Environmental Health Sciences

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# Integrating population genomic data to understand mechanisms of chemical susceptibility and resistance

Boston Univ. and Duke Univ. SRPs

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Consultants/Collaborators: Andrew Whitehead (UC Davis); Wes Warren (U Missouri), Diane Nacci and Bryan Clark (U.S. EPA, Office of Research and Development)









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# **Research Questions**

- What are the impacts of long-term exposure to toxic chemicals on natural populations?
- Can populations adapt over time? What are the mechanisms of adaptation? What are the fitness costs of adaptation?
- How does genetic variation influence sensitivity to toxic chemicals (GxE)?

# **Population-based Model System**

 Atlantic killifish (*Fundulus heteroclitus*) inhabiting estuaries (including Superfund sites) along the U.S. East Coast.







Atlantic killifish (Fundulus heteroclitus) (photo courtesy of Dr. Evan D'Alessandro, RSMAS)

# Goals

- <u>Integrate</u> two large genome-sequencing data sets (and related data) from two SRP Centers that both study the genetic mechanisms underlying evolved resistance to
  - PCBs and dioxins (Boston University)
  - PAHs (Duke University)

in Atlantic killifish populations living at Superfund sites.

- <u>Enhance</u> the findability, accessibility, interoperability, and reuse (FAIR) of the data sets.
- <u>Enable</u> comparison of these datasets to other data (zebrafish, mouse, human) on genetic variation associated with chemical pollution and disease.



#### A. **DL-PCBs** New Bedford Harbor, MA Τ2 T1 **S1** T3 Bridgeport, CT **DL-PCBs**, PAHs S2 AtlanticOcean **S**3 **Chlorinated dioxins** Newark, NJ **S**4 **PAHs** Elizabeth River, VA T4 USA T=Tolerant S=Sensitive (DL-PCBs = dioxin-like PCBs)

# Tolerant (T) and Sensitive (S) populations of killifish: Whole-genome sequence data (Boston Univ SRP and collaborators)

Reid et al. 2016 Science 354: 1305

## Tolerant and Sensitive populations of killifish: RAD-seq data (Duke SRP)





Osterberg *et al.* 2018 *Aquat Toxicol* **200**: 73, and unpublished

## Populations vary in embryo sensitivity to PCB-126 (up to 8,000-fold)



Genome-wide approaches to understand the mechanism of resistance

- Gene expression profiling (Whitehead et al. (2012) Proc Royal Soc Biol Sci 279: 427-433)
- Quantitative genetics [quantitative trait loci (QTL)] (Nacci et al. (2016) Mol Ecol 25: 5467-5482)
- Restriction-site Associated DNA (RAD)-sequencing (RAD-seq) (Osterberg et al. (2018) Aquat Toxicol 200: 73-82)
- Whole Genome Sequencing (WGS) (Reid *et al.* (2016) *Science* **354**: 1305-1308)

### Aryl Hydrocarbon Receptor (AHR) pathway as a Shared Target of Selection



cyp1a, cyp1b, gst, ahrr,...

## **Existing data sets**

- <u>Whole-genome re-sequencing (WGS) data</u> for 384 individual killifish in eight populations (four pairs of tolerant and sensitive populations) [Reid, et al. (2016) Science 354: 1305-1308].
- <u>RAD-seq data</u> (genome sequence samples) from 288 individual fish from nine populations [Osterberg, *et al.* (2018) *Aquat Toxicol* **200**: 73-82].
- <u>RNA-seq</u> data.
- *F. grandis* <u>WGS data</u> from nine populations [Oziolor, et al. (2019) *Science* **364**: 455-457].
- Fundulus heteroclitus <u>New genome assembly</u> (NCBI) [Warren, Whitehead, et al. (2020) PRJNA615222]

#### Number of data sets

WGS	RAD-seq	RNA-seq	TOTAL
672	288	75	1035

# Challenges

- How to integrate these disparate data sets (WGS, RADseq, RNA-seq) to allow analysis and reuse
- How to retain access to population, individual, and molecular-level information on genetic variants
- How to make the data easily accessible, expandable, and usable by other researchers
- How to enable comparison of these data with GxE data in other species









# **Pipeline**

WGS	RAD-seq	RNA-seq	TOTAL
672	288	75	1035

- Harmonized bioinformatics analysis pipelines
- Reanalyzed data using new *Fundulus heteroclitus* reference genome assembly
- Loaded data and metadata into the opensource genome browser JBrowse [Buels et al. (2016) Genome Biology 17:66]
- Performed joint variant calling

SuperFunBase

http://fundulus.bu.edu



## SuperFunBase: JBrowse Interface



# SuperFunBase: Joint variant (SNV) calling



# SuperFunBase: Population-specific variants

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## SuperFunBase: Larger genomic features



# Open Science Framework (OSF.io)



# What's next?

- Add population-specific epigenomic data (esp. DNA methylation)
- Call SNVs from RNA-seq data
- Strategy for linking killifish SNV data with SNV data for orthologous genes in other species (humans, rodents, zebrafish)

Example: AIP (AHR-Interacting Protein)



# AIP mutations predispose to Familial Isolated Pituitary Adenoma (FIPA)



https://www.qmul.ac.uk/fipa-patients/



Comparing SNVs in Orthologous Genes Across Populations and Species

# **Other Comparative Databases**

• Comparative Toxicogenomics Database (CTD)

"Chemical–gene/protein interactions, chemical–disease and gene–disease relationships" http://ctdbase.org

 SeqAPASS (Sequence Alignment to Predict Across Species Susceptibility) "Predict across-species susceptibility to chemicals with known molecular targets"

https://www.epa.gov/chemical-research/sequence-alignment-predictacross-species-susceptibility

 Monarch Initiative of the Global Alliance for Genomics and Health (GA4GH) "Connecting phenotypes to genotypes across species" <u>https://monarchinitiative.org</u> Integrating population genomic data to understand mechanisms of chemical susceptibility and resistance



SuperFunBase

http://fundulus.bu.edu

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