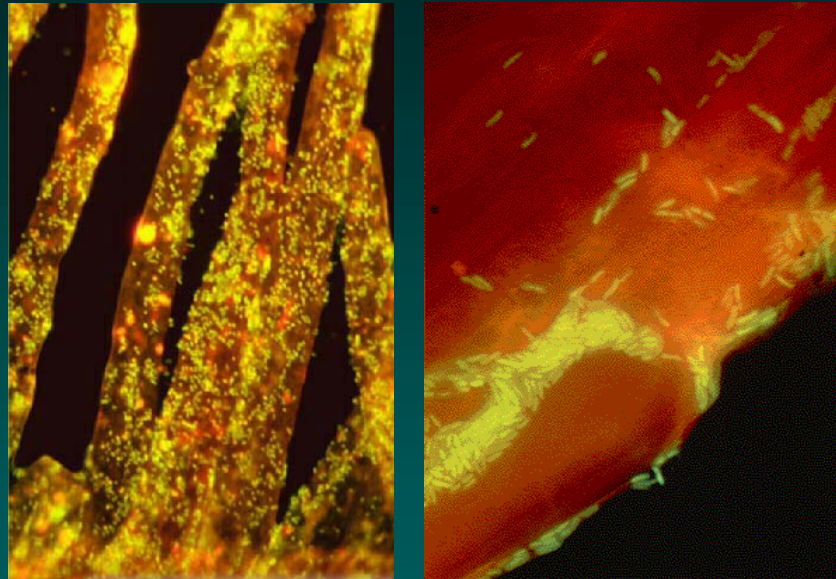


Phytoremediation of PCBs and PAHs with Monoterpene Producing Plants



David Crowley, Joong-Wook Park, Haakrho Yi
Department of Environmental Sciences
University of California, Riverside

Xenobiotic



Plant

Uptake
Metabolism
Volatilization
Carbon
exudate
quantity
components
Surfactants
O₂, CO₂, pH
Nitrogen



Rhizosphere Community

Species Composition
Population numbers
Degradars
Cometabolizers
Activity
Catabolite repression
Gene recombination
Plasmid transfer



Soil

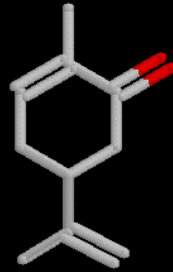
Solubility
Adsorption
% OM
Clay
Organic matter
Texture
water content
aeration
Temperature
Nutrient Content



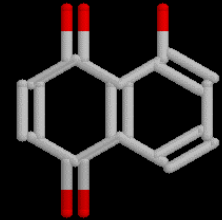
Metabolite Formation
Mineralization

Phytochemical Analogs of Xenobiotic Contaminants

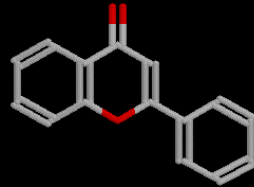
Terpenes:



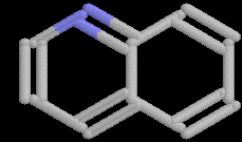
Quinones



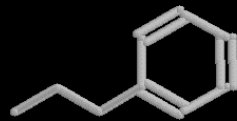
Flavonoids



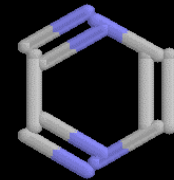
Alkaloids



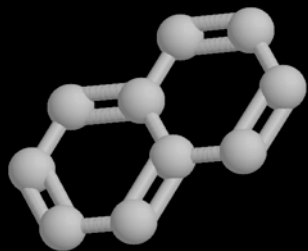
Lignin



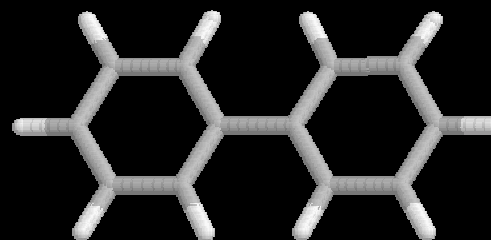
Pyrazines



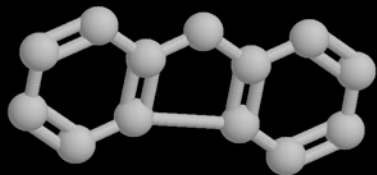
Structures of Common Aromatic Soil Pollutants



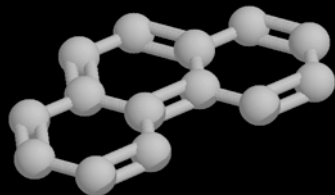
Napthalene



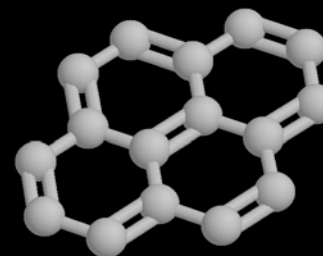
Polychlorinated Biphenyls



Fluorene

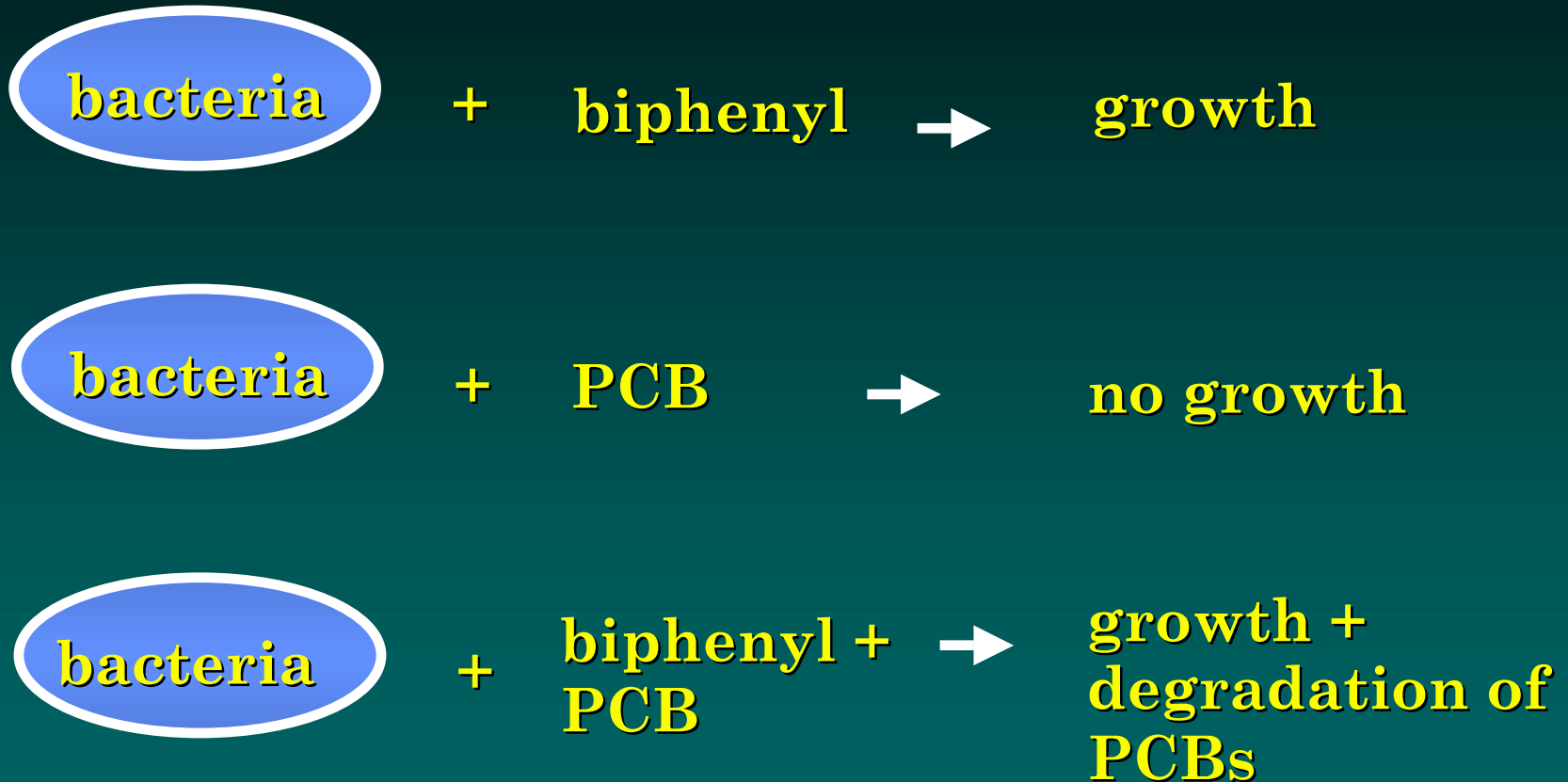


Phenanthrene

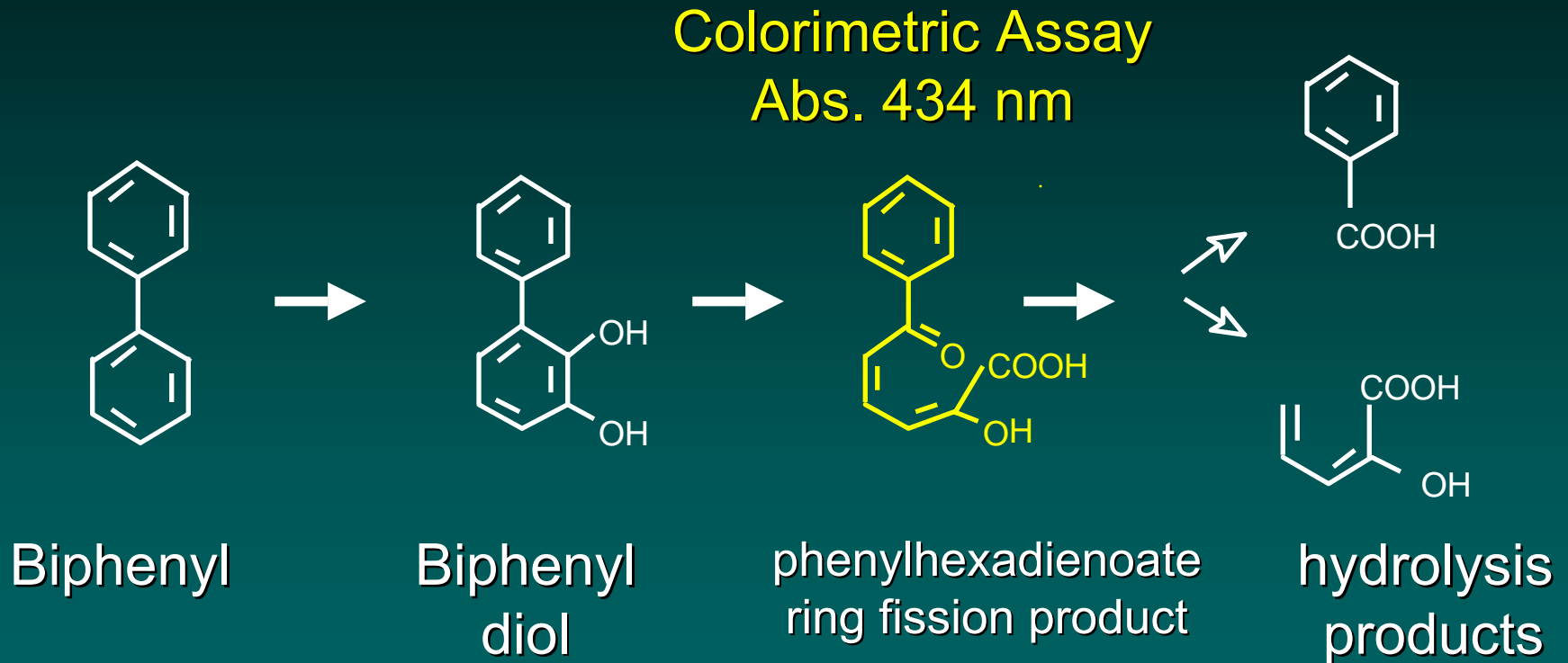


Pyrene

Cometabolism of PCBs



Assay for Cometabolism of PCBs

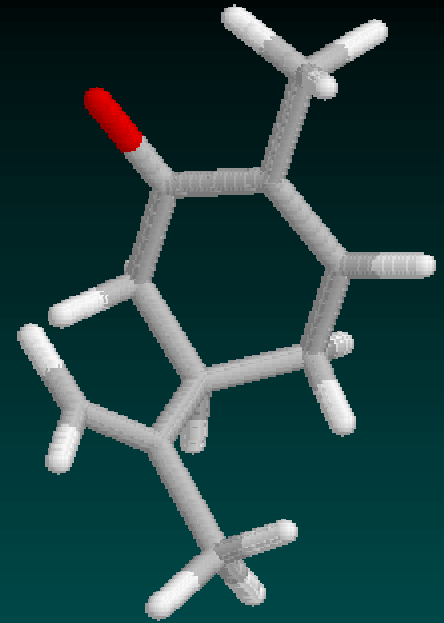


Plant Screening Assay

Gilbert and Crowley 1996

Spearmint (*Mentha spicata*)

Active component: carvone

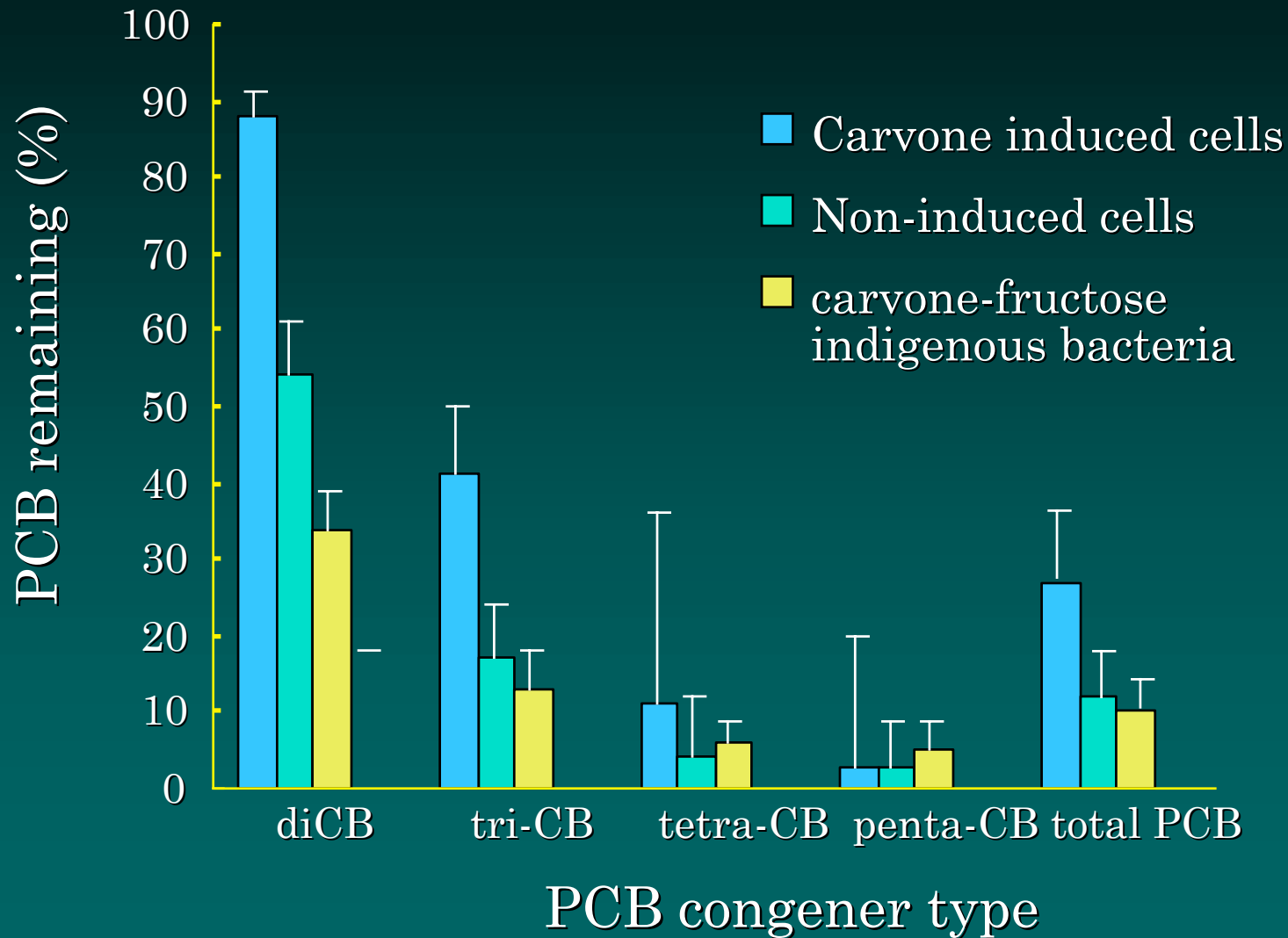


Arthrobacter B1B

Evidence:

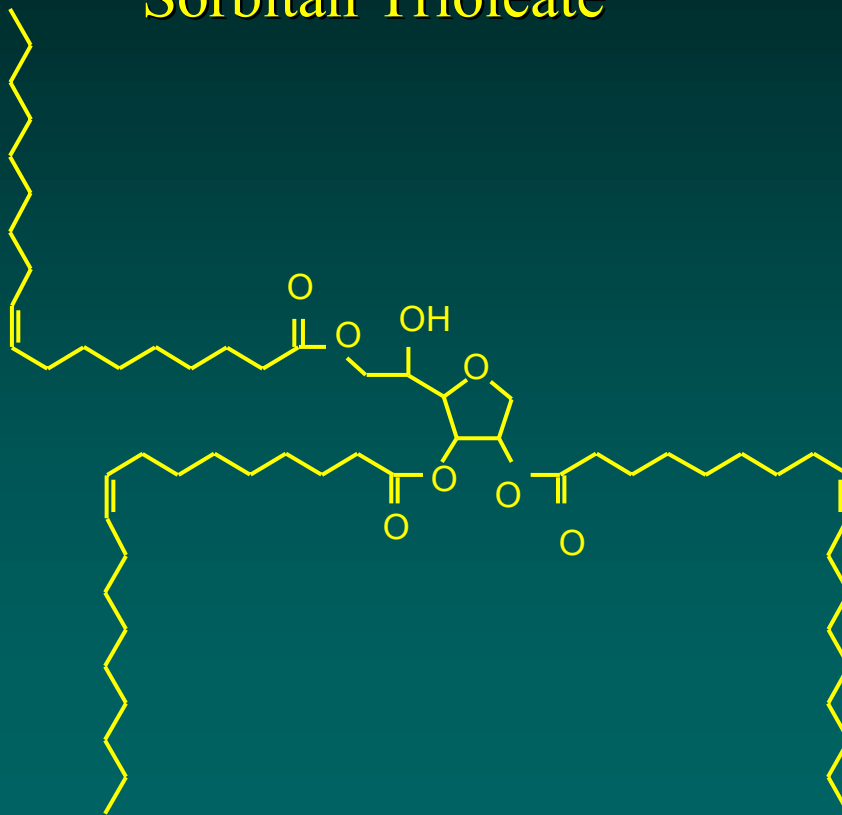
PCB ring oxidation product
chlorobenzoate product formation
disappearance of Aroclor 1242

Degradation of 100 ppm Aroclor 1242 PCBs in Soil after 9 Weeks After Inoculation With Carvone-Induced *Arthrobacter* B1B



Development of a Field Application Vector

Sorbitan Trioleate



Criteria:

Nontoxic

Selective Growth Substrate

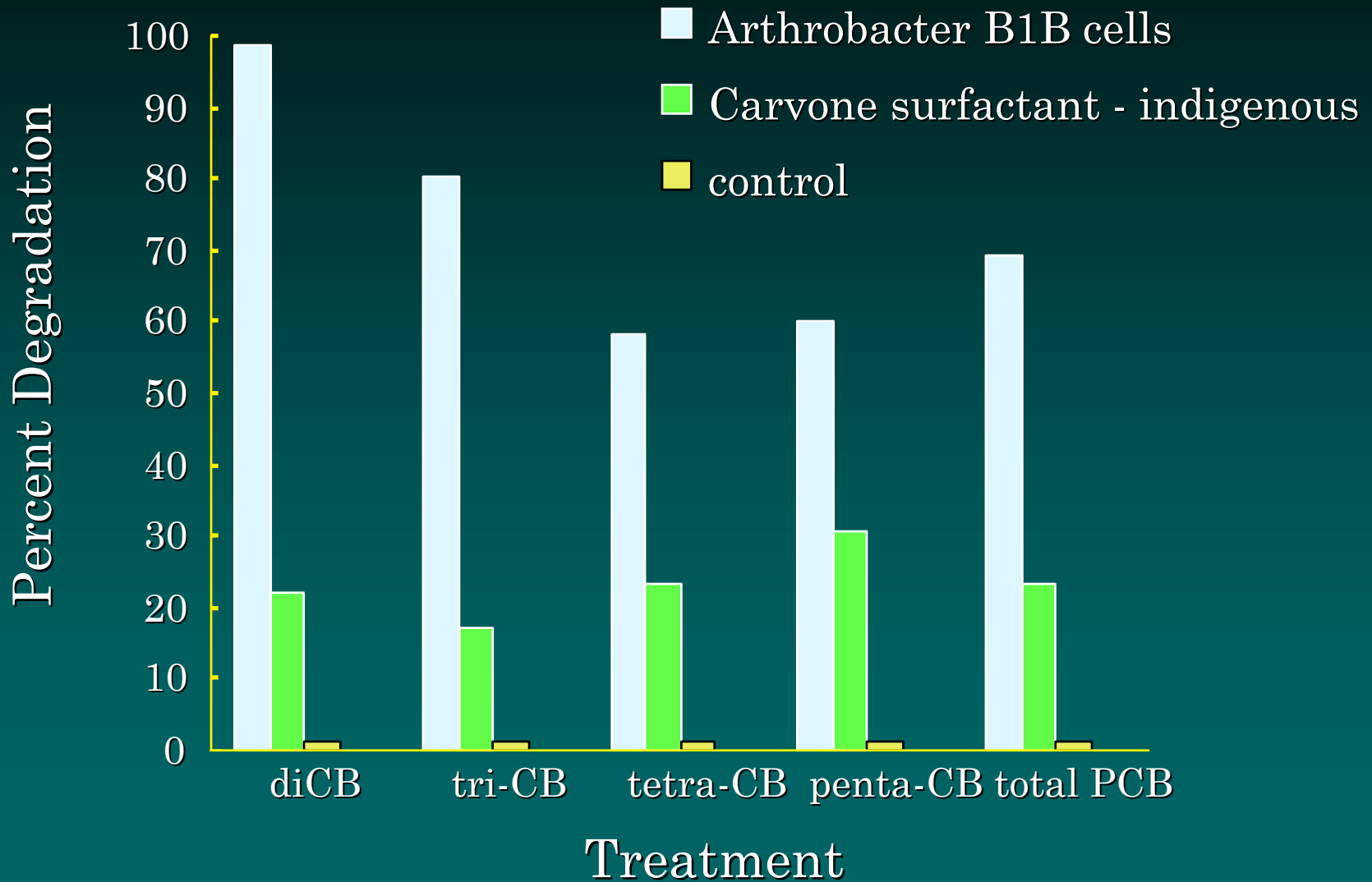
Solubilizes PCB

Biodegradable

Fatty Acid Composition:

Oleic acid	74%	
Linoleic acid	7%	7%
Linolenic acid	2%	
Palmitoleic acid	7%	
Palmitic acid	10%	

Degradation of Aroclor 1242 PCBs after 9 Weeks of Repeated Inoculation Using Carvone-Surfactant Grown Cells



Induction of PCB degradation by Carvone and Related Compounds

TABLE 3. Ring fission product formation induced by carvone and structurally similar compounds

Compound ^a	Product formation rate ^b	Significance ^c	Plant source ^d
(1) <i>p</i> -Cymene	1,471 ± 343	a	Widely distributed
(2) Isoprene	928 ± 52	b	Widely distributed
(3) (<i>S</i>)-(+)-Carvone	860 ± 117	bc	Dill seed and caraway seed
(4) (<i>R</i>)-(-)-Carvone	845 ± 187	bc	Spearmint
(5) (<i>S</i>)-(-)-Limonene	821 ± 168	bc	Pine needle oil
(6) (<i>R</i>)-(+)-Limonene	807 ± 103	bc	Citrus, juniper, and dill seed
(7) Biphenyl	706 ± 72	c	None
(8) Carvacrol	509 ± 19	d	Oregano
(9) Cumene	280 ± 110	e	None
(10) <i>p</i> -Xylene	188 ± 80	ef	None
(11) Toluene	138 ± 23	ef	None
(12) <i>trans</i> -Cinnamic acid	87 ± 3	f	Storax
(13) Thymol	70 ± 10	f	Thyme
(14) Benzene	61 ± 13	f	None
Fructose only (control)	73 ± 48	f	NA ^e

^a Numbers correspond to those for structures in Fig. 2.

^b Values are nanograms of product per milligram of protein per hour.

^c Rates of compounds marked by same letter are not significantly different ($P < 0.05$; statistical analysis by Fisher's least significant difference test).

^d Major plant source according to Buckingham (5).

^e NA, not applicable.

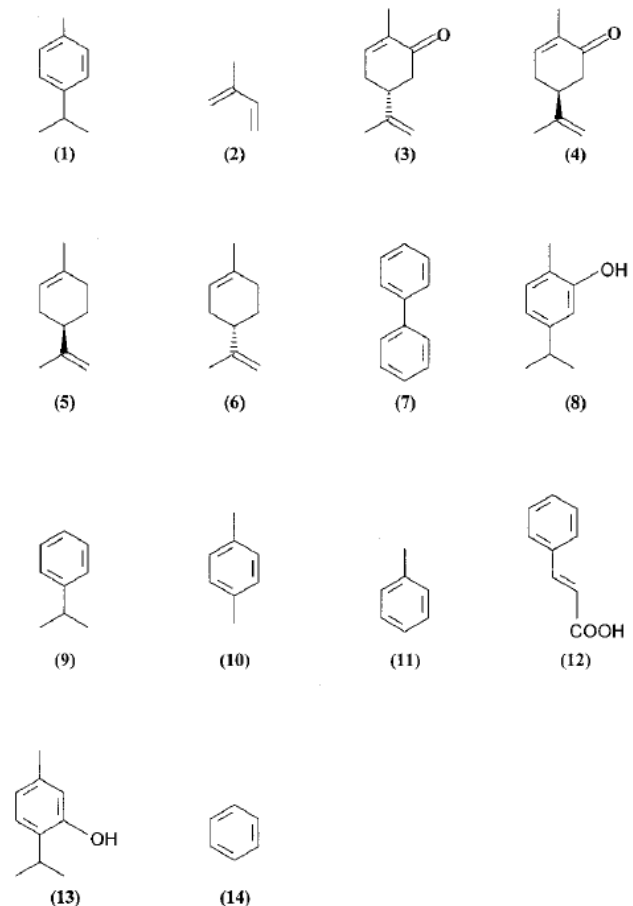
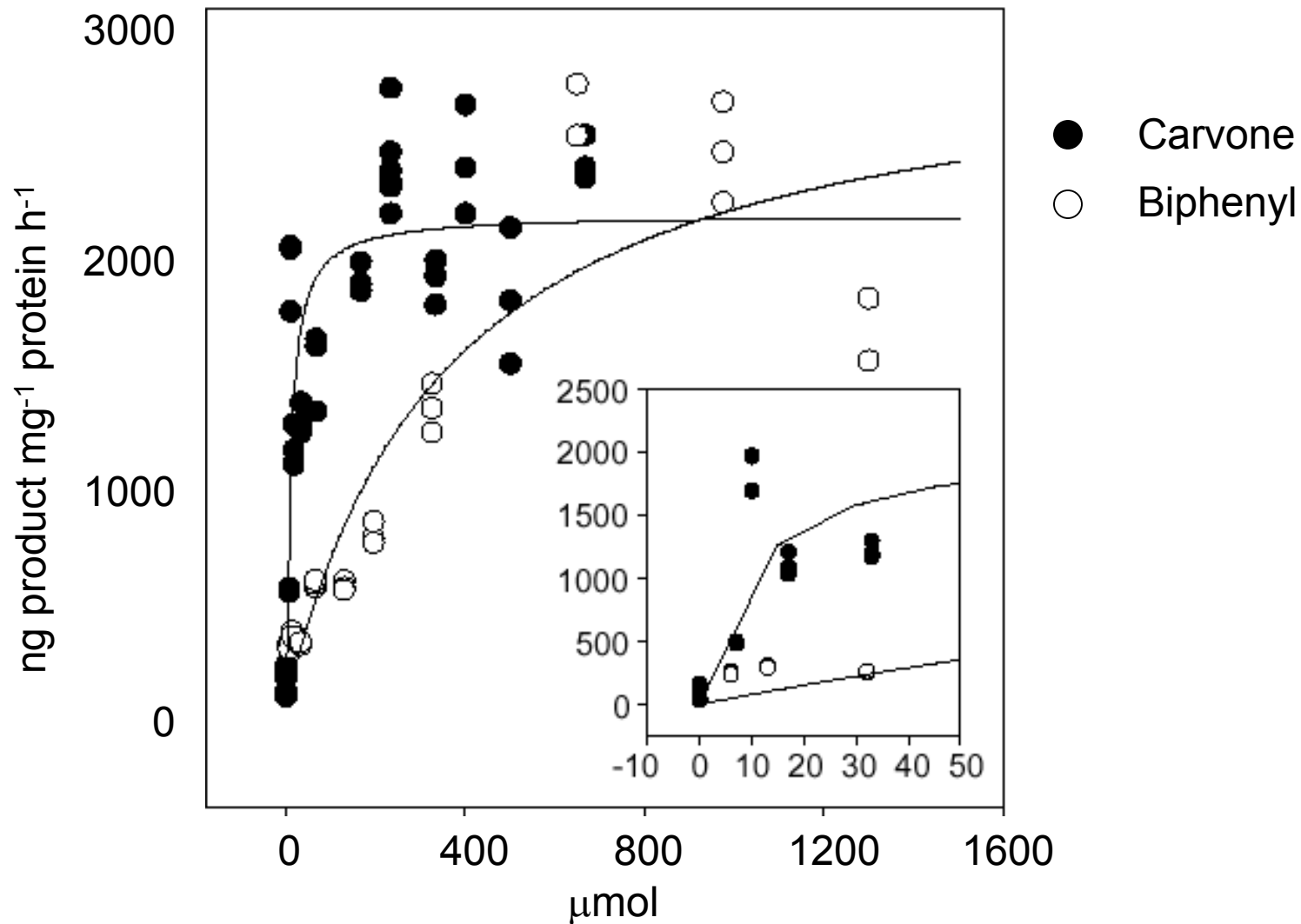
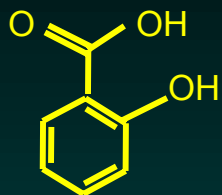


FIG. 2. Structures of compounds used in this study.

PCB ring fission product formation by *Arthrobacter* B1B following enzyme induction by carvone or biphenyl. (Gilbert and Crowley 1998)

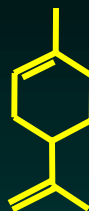




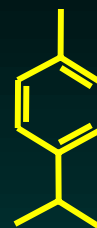
Salicylate



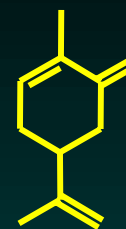
cumene



limonene



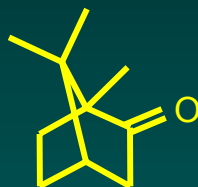
cymene



carvone



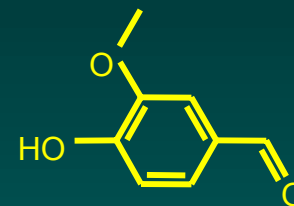
cineole



camphor



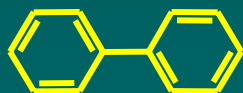
pinene



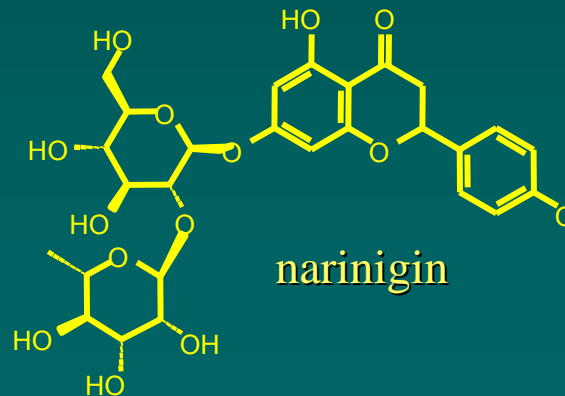
vanillin



citral



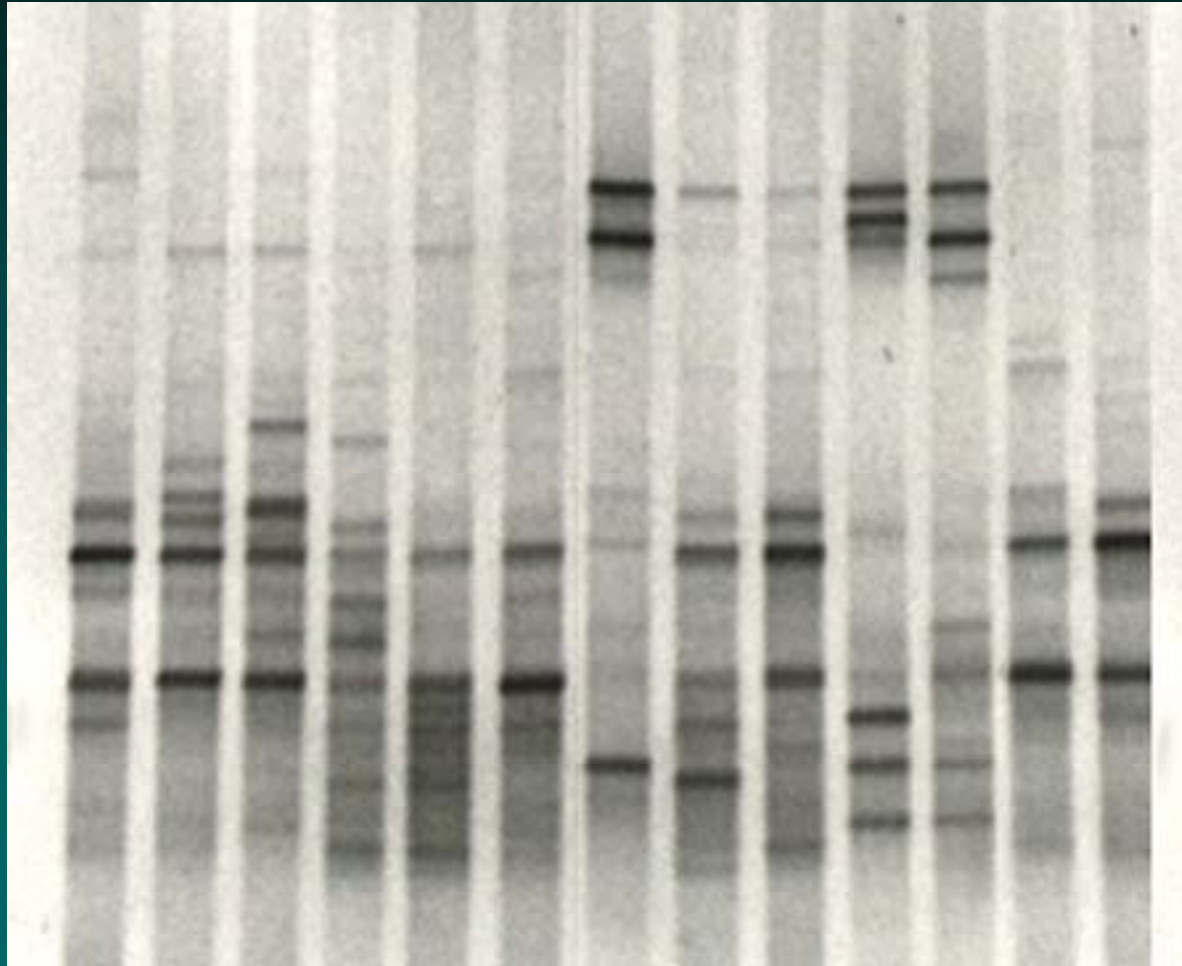
biphenyl



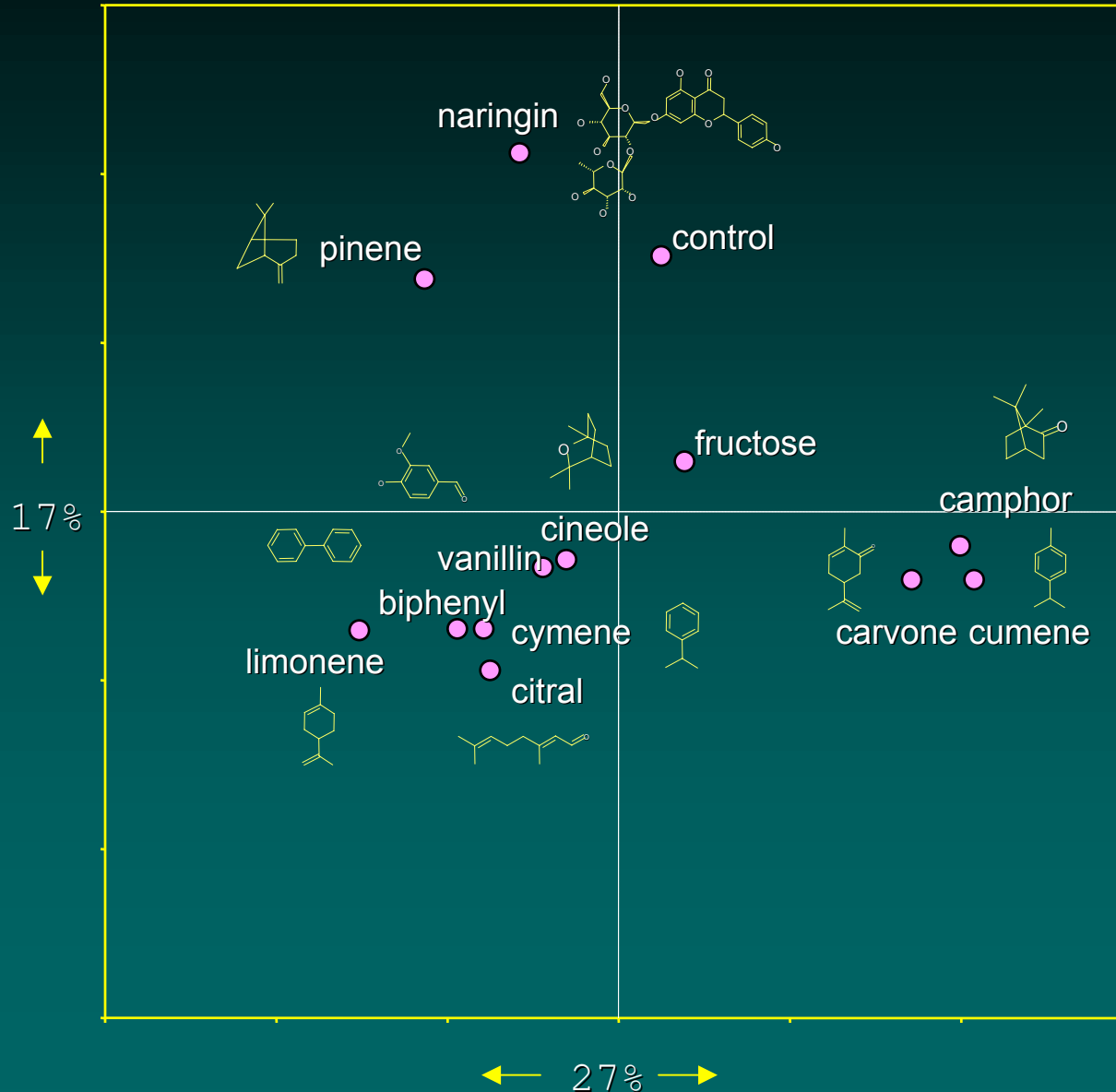
naringin

BRDU Labeled 16S rDNA Profile of Terpene Enriched Soil

Biphenyl
Vanillin
Citral
Pinene
Limonene
Naringin
Cumene
Cymene
Cineole
Carvone
Camphor
Fructose
Control



Correspondence Analysis of BRDU Labeled 16S rDNA from Terpene Enriched Soils



Salicylate: Plant Signal Compound, Siderophore, and Inducer of Xenobiotic Degradation Enzymes

Toluene dioxygenase substrates:

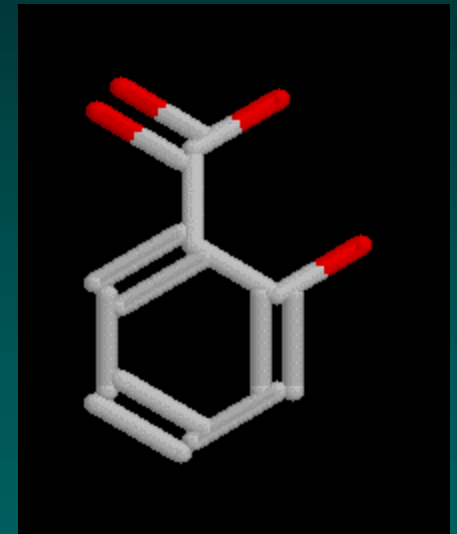
Benzene, toluene, xylene (BTEX)

Trichloroethylene (TCE)

Trinitrotoluene (TNT)

Naphthalene, benzopyrene (PAH)

Polychlorinated biphenyls (PCB)



Coinoculation with *Arthrobacter* sp. B1B and *Ralstonia eutrophus* H850

PCB congener	<i>Arthrobacter</i>	<i>Ralstonia</i> coinoculation
2,3,6,2'	34 ± 5	45 ± 6
2,5,2',5'	29 ± 3	56 ± 7
2,4,2',5'	27 ± 3	49 ± 6
2,4,2',4', 2,4,5,2'	30 ± 5	44 ± 6
2,3,2',3'	30 ± 6	51 ± 6
2,3,5,2',4', 2,4,5,2',5	13 ± 4	25 ± 4

Summary

- Gram positive bacteria induced by monoterpenes.
- Gram negative bacteria induced by salicylic acid.
- Many indigenous PCB degraders can be enriched by growth on these substances.

Common terpene producing plant species

Origanum majorana

O. vulgare

Thymus zygis

Curcuma longa

Eucalyptus species

Vitex negundo

Ligusticum porteri

Andropogon sp.

O. syriacum

Salvia cabulica

T. piperella

Carum copticum

Tanacetum parthenium

Acanthopanax trifoliatum

Ocimum basilicum

Chenopodium sp.

Chenopodium ambrosioides



Alpha-pinene, Aritasone, Ascaridole, Ascorbic-acid, Beta-carotene, Butyric-acid, Calcium, D-camphor, EO, Ferulic-acid, Geraniol, L-pinocarvone, Leucine, Limonene, Malic-acid, Menthadiene, Methyl-salicylate, Myrcene, Niacin, P-cymene, P-cymol, Phosphorus, Safrole, Saponins, Spinasterol, Tartaric-acid, Terpinene, Terpinyl-acetate, Terpinyl-salicylate, Thiamin, Triacontyl-alcohol, Trimethylamine, Urease, Vanillic-acid



Phytoremediation of Pyrene using Celery Root

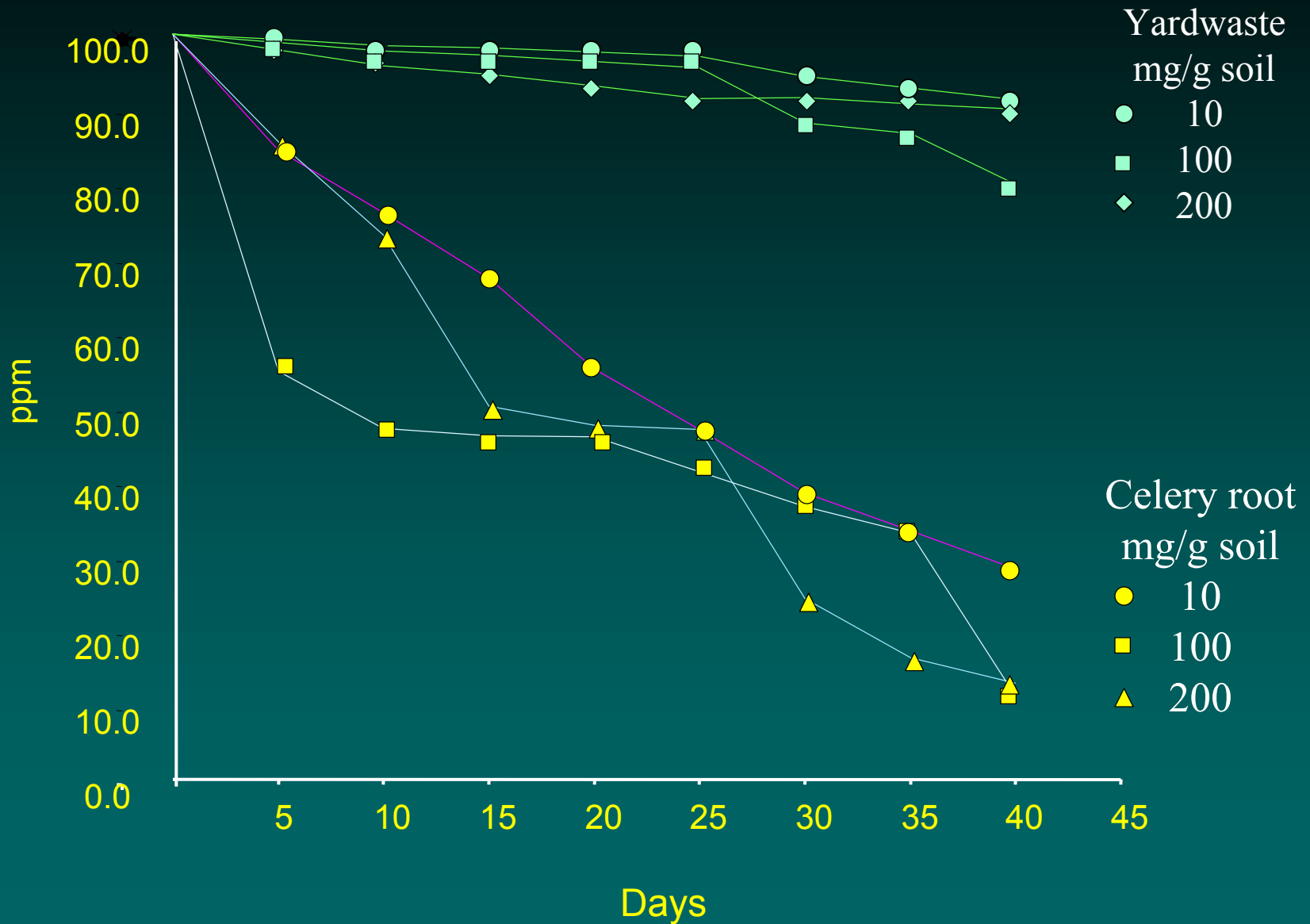


Root essential oil:

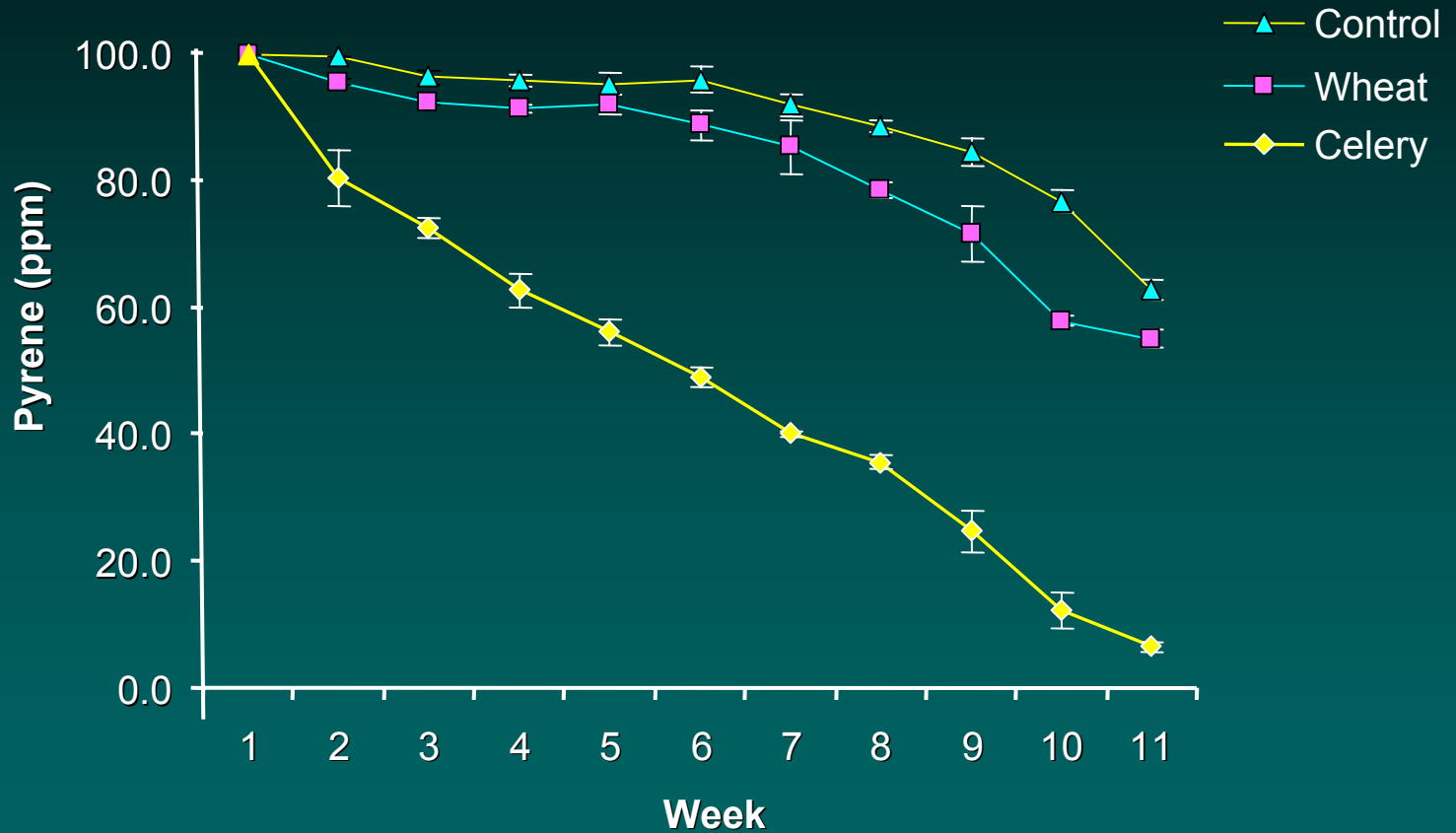
	<u>ppm</u>
β pinene	15,000
carvone	5000
dihydro-carvone	5000
P-cymene	31,000
limonene	117,000
myrcene	18,000
terpenoline	33,000
trans-ocimene	290,000
cis-ocimene	68,000

Duke, J. A. 1992. Handbook of phytochemical constituents of GRAS herbs and other economic plants. Boca Raton, FL. CRC Press.

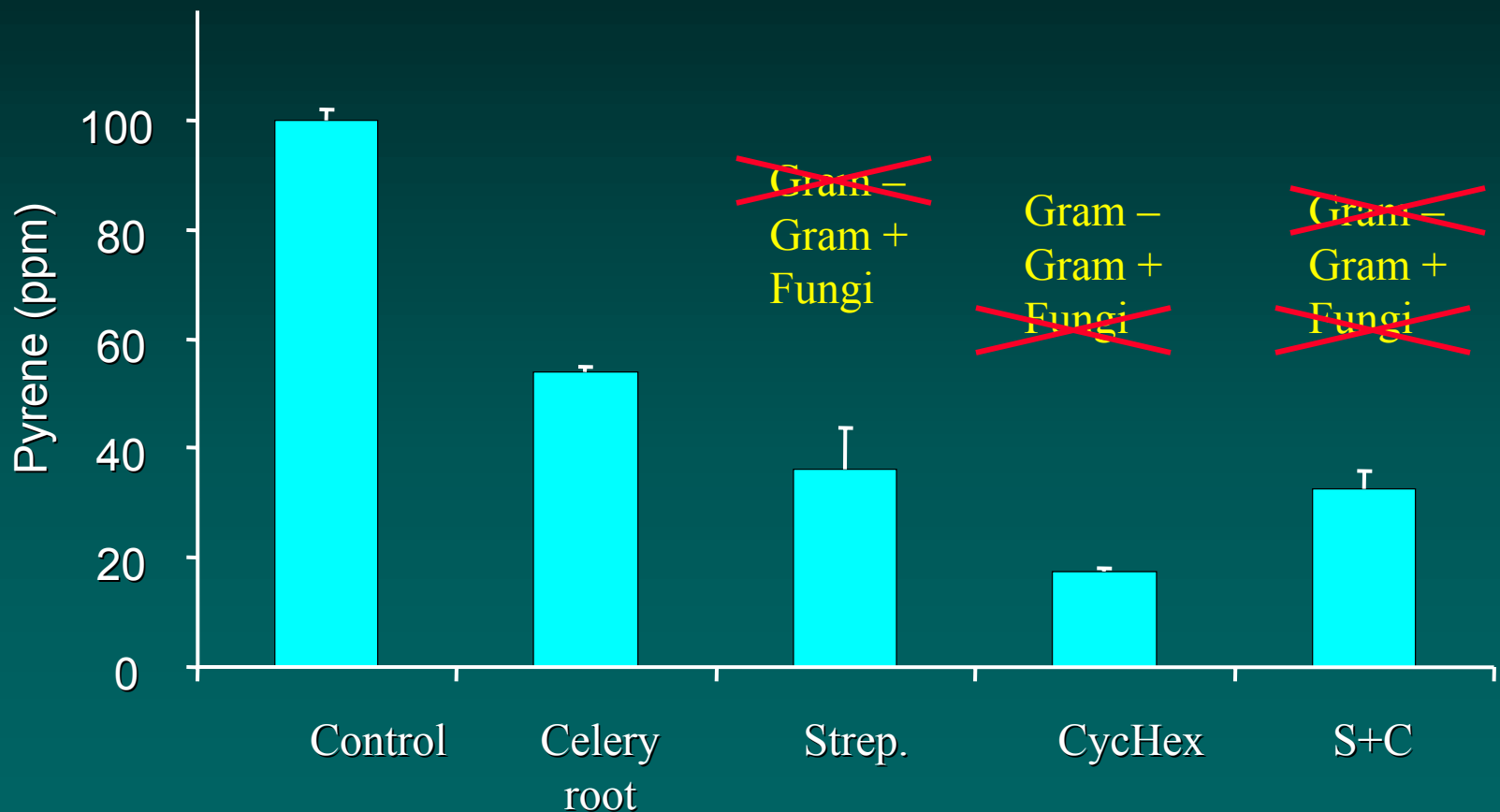
Degradation of pyrene in soil amended with celery root



Degradation of pyrene in the rhizosphere of wheat and celery plants

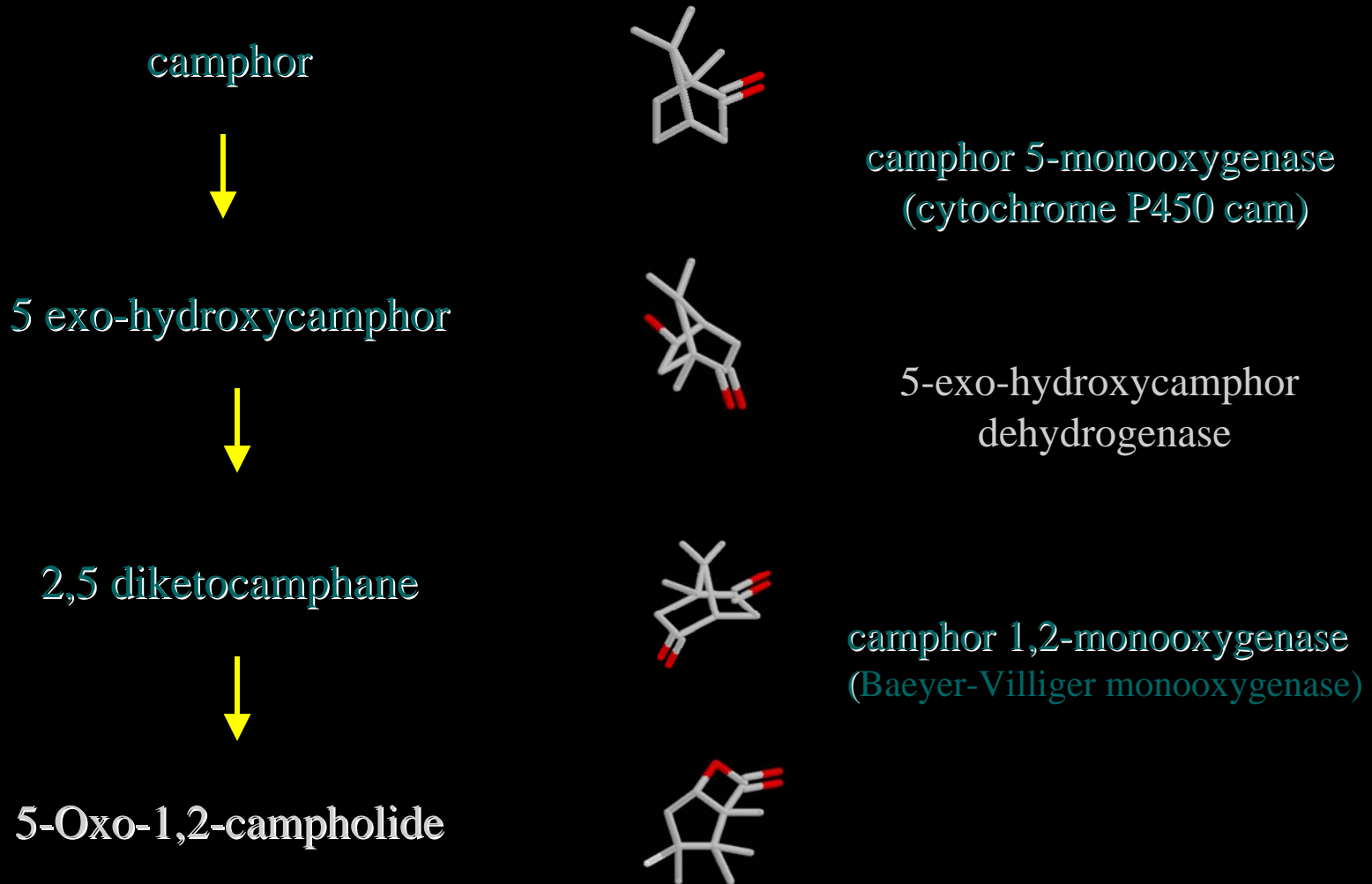


Selective inhibitors suggest Gram + bacteria are predominant pyrene degraders induced by celery root.



What enzymes are induced by monoterpenes?

Biodegradation of Camphor: Role of P450CAM



Amino Acid Sequence Alignments of Riske Iron Center Regions of Group 3 PAH and Terpene Dioxygenases

DitA	96	VTLNVOPHRGMRISTADCGNTQ-IHKOIYHGWAFR	119...201	IVNANWKTAGEQSAADGFHT-LTLHRWLGE	229
NidA	83	GHLNAORHRGMQVCRAEMGNAS-HFROPYHGWTYS	116...198	VVDANWKLGADNFVGDAYHT-MMTHRSMVE	226
NahAc G4	76	AFLNVORHRGKTLVNAEAGNAK-GFVOSYHGWGFG	109...190	VIKANWKAPAENFVGDAYHV-GWTHASSLR	218
PahAc	76	AFLNVORHRGKTLVNAEAGNAK-GFVOSYHGWGFG	109...190	VIKANWKAPAENFVGDAYHV-GWTHASSLC	218
DoxB	76	AFLNVORHRGKTLVSVEAGNAK-GFVOSYHGWGFG	109...190	VIKANWKAPAENFVGDAYHV-GWTHASSLR	218
NdoB	76	AFLNVORHRGKTLVSVEAGNAK-GFVOSYHGWGFG	109...190	VIKANWKAPAENFVGDAYHV-GWTHASSLR	218
NahAc 9816-4	76	AFLNVORHRGKTLVSVEAGNAK-GFVOSYHGWGFG	109...190	VIKANWKAPAENFVGDAYHV-GWTHASSLR	218
PahA3	76	AFLNVORHRGKTLVHAEAGNAK-GFVOSYHGWGFG	109...190	IIKANWKAPAENFTGDAYHV-GWTHASSLR	218
NtdAc	74	AFLNVORHRGKTLVHTEAGNAK-GFVOGYHGWGYG	107...188	VVKANWKPFAENFVGDYHV-GWTHAAALR	216
DntAc	79	AFLNVORHRGKTIVDAEAGNAK-GFVOGYHGWGYG	112...193	VVKGNWKVFAENFVGDYHI-GWTHASILR	221
NagAc	74	AFLNVORHRGKTLVHAEAGNAK-GFVOSYHGWGFG	107...188	VIKANWKAPAENFVGDAYHV-GWTHASSLR	216
PhnAc RP007	77	AFLNVORHRGARLCAVEAGNAR-GFAONYHGWAYG	110...191	FIEANWKAPSENFVGDAYHV-GWTHASALR	219
PhnAc AFK2	81	AFLNVOTHRGARLVAAEAANAR-AFSOTYHGWSFG	114...194	LLNCNWKTPAENFVGDAYHV-GWTHLASLM	222

Conclusions

- Degradation of xenobiotics in the rhizosphere is likely enhanced by a combination of mechanisms, including: fortuitous enrichment of degraders, growth linked metabolism, and cometabolism.
- Plant signal molecules including terpenes, flavonoids, and salicylate have multiple ecological functions. When released into the rhizosphere, these compounds also have signal functions for inducing cometabolism of organic xenobiotics.

Research Needs

- Identification of specific plants that produce substances that can biostimulate xenobiotic degradation.
- Evaluation of the role of plant, microbial, and earthworm enzymes (oxygenase, cytochrome P450 ...) for effectiveness in degrading soil contaminants.
- Studies on the ecology of rhizosphere microbial consortia for enhancing bioremediation.

Acknowledgments

Eric Gilbert

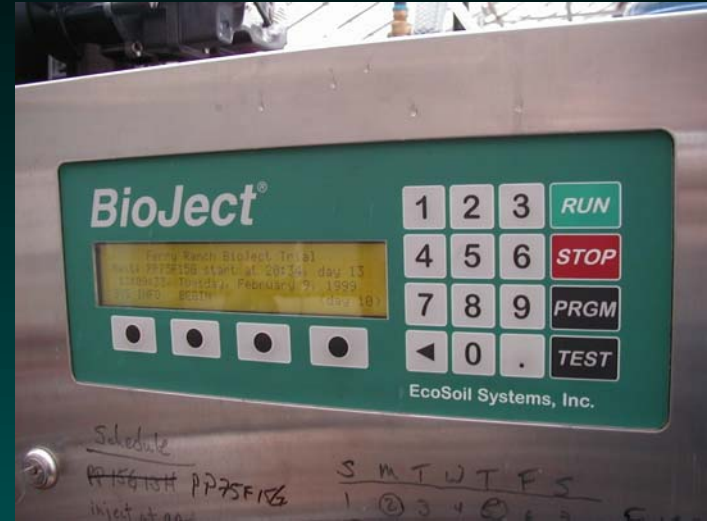
Andrew Singer

Ekawan Luepromchai

Joong Wook Park

Haakrho Yi

Development of Methods for Soil Bioaugmentation



Enzymes for Transformations of Monoterpenes

